

Introduction to RBM package

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1 Overview

This document provides an introduction to the `RBM` package. The `RBM` package executes the resampling-based empirical Bayes approach using either permutation or bootstrap tests based on moderated t-statistics through the following steps.

- Firstly, the `RBM` package computes the moderated t-statistics based on the observed data set for each feature using the `lmFit` and `eBayes` function.
- Secondly, the original data are permuted or bootstrapped in a way that matches the null hypothesis to generate permuted or bootstrapped resamples, and the reference distribution is constructed using the resampled moderated t-statistics calculated from permutation or bootstrap resamples.
- Finally, the p-values from permutation or bootstrap tests are calculated based on the proportion of the permuted or bootstrapped moderated t-statistics that are as extreme as, or more extreme than, the observed moderated t-statistics.

Additional detailed information regarding resampling-based empirical Bayes approach can be found elsewhere (Li et al., 2013).

2 Getting started

The RBM package can be installed and loaded through the following R code.
Install the RBM package with:

```
> if (!requireNamespace("BiocManager", quietly=TRUE))
+   install.packages("BiocManager")
> BiocManager::install("RBM")
```

Load the RBM package with:

```
> library(RBM)
```

3 RBM_T and RBM_F functions

There are two functions in the RBM package: `RBM_T` and `RBM_F`. Both functions require input data in the matrix format with rows denoting features and columns denoting samples. `RBM_T` is used for two-group comparisons such as study designs with a treatment group and a control group. `RBM_F` can be used for more complex study designs such as more than two groups or time-course studies. Both functions need a vector for group notation, i.e., "1" denotes the treatment group and "0" denotes the control group. For the `RBM_F` function, a contrast vector need to be provided by users to perform pairwise comparisons between groups. For example, if the design has three groups (0, 1, 2), the `aContrast` parameter will be a vector such as ("X1-X0", "X2-X1", "X2-X0") to denote all pairwise comparisons. Users just need to add an extra "X" before the group labels to do the contrasts.

- Examples using the `RBM_T` function: `normdata` simulates a standardized gene expression data and `unifdata` simulates a methylation microarray data. The p -values from the `RBM_T` function could be further adjusted using the `p.adjust` function in the `stats` package through the Benjamini-Hochberg method.

```
> library(RBM)
> normdata <- matrix(rnorm(1000*6, 0, 1),1000,6)
> mydesign <- c(0,0,0,1,1,1)
> myresult <- RBM_T(normdata,mydesign,100,0.05)
> summary(myresult)
```

	Length	Class	Mode
ordfit_t	1000	-none-	numeric
ordfit_pvalue	1000	-none-	numeric
ordfit_beta0	1000	-none-	numeric
ordfit_beta1	1000	-none-	numeric
permutation_p	1000	-none-	numeric
bootstrap_p	1000	-none-	numeric

```
> sum(myresult$permutation_p<=0.05)
```

```

[1] 24

> which(myresult$permutation_p<=0.05)

[1] 1 59 88 100 104 113 170 197 251 256 260 322 368 396 492
[16] 505 593 686 793 839 850 908 990 1000

> sum(myresult$bootstrap_p<=0.05)

[1] 5

> which(myresult$bootstrap_p<=0.05)

[1] 104 152 321 425 700

> permutation_adj_p <- p.adjust(myresult$permutation_p, "BH")
> sum(permutation_adj_p<=0.05)

[1] 1

> bootstrap_adj_p <- p.adjust(myresult$bootstrap_p, "BH")
> sum(bootstrap_adj_p<=0.05)

[1] 0

> unifdata <- matrix(runif(1000*7,0.10, 0.95), 1000, 7)
> mydesign2 <- c(0,0,0, 1,1,1,1)
> myresult2 <- RBM_T(unifdata,mydesign2,100,0.05)
> sum(myresult2$permutatioin_p<=0.05)

[1] 0

> sum(myresult2$bootstrap_p<=0.05)

[1] 38

> which(myresult2$bootstrap_p<=0.05)

[1] 3 11 12 52 106 141 159 189 202 230 273 276 299 303 321 334 349 358 385
[20] 411 429 432 473 512 534 563 587 590 610 639 685 719 756 822 884 951 953 983

> bootstrap2_adj_p <- p.adjust(myresult2$bootstrap_p, "BH")
> sum(bootstrap2_adj_p<=0.05)

[1] 0

```

- Examples using the RBM_F function: normdata_F simulates a standardized gene expression data and unifdata_F simulates a methylation microarray data. In both examples, we were interested in pairwise comparisons.

```

> normdata_F <- matrix(rnorm(1000*9,0,2), 1000, 9)
> mydesign_F <- c(0, 0, 0, 1, 1, 1, 2, 2, 2)
> aContrast <- c("X1-X0", "X2-X1", "X2-X0")
> myresult_F <- RBM_F(normdata_F, mydesign_F, aContrast, 100, 0.05)
> summary(myresult_F)

              Length Class  Mode
ordfit_t      3000   -none-  numeric
ordfit_pvalue 3000   -none-  numeric
ordfit_beta1   3000   -none-  numeric
permutation_p 3000   -none-  numeric
bootstrap_p    3000   -none-  numeric

> sum(myresult_F$permutation_p[, 1]<=0.05)

[1] 71

> sum(myresult_F$permutation_p[, 2]<=0.05)

[1] 73

> sum(myresult_F$permutation_p[, 3]<=0.05)

[1] 70

> which(myresult_F$permutation_p[, 1]<=0.05)

[1] 10 16 51 63 64 81 88 92 98 99 101 102 132 136 144 147 156 187 213
[20] 214 223 242 246 250 255 259 294 295 304 314 324 326 335 349 350 423 426 448
[39] 468 476 488 520 539 566 574 580 582 607 619 636 653 656 657 670 674 693 725
[58] 802 825 831 837 868 881 889 897 898 911 932 953 979 992

> which(myresult_F$permutation_p[, 2]<=0.05)

[1] 16 51 63 64 81 88 92 98 99 101 102 132 144 147 156 187 213 214 219
[20] 223 242 259 294 295 304 314 324 326 346 349 350 423 426 468 476 488 520 539
[39] 566 574 580 591 607 610 619 636 638 653 656 657 670 674 693 699 702 707 712
[58] 725 802 831 834 837 868 881 889 897 898 901 908 911 932 953 992

> which(myresult_F$permutation_p[, 3]<=0.05)

[1] 10 16 51 64 81 88 90 92 98 101 102 132 136 144 147 187 194 214 219
[20] 223 242 250 259 287 294 295 304 324 335 349 423 426 448 460 468 476 488 504
[39] 520 539 566 574 580 607 619 636 653 656 657 668 670 674 693 707 725 802 825
[58] 826 831 837 868 881 897 898 901 911 932 970 979 992

> con1_adjp <- p.adjust(myresult_F$permutation_p[, 1], "BH")
> sum(con1_adjp<=0.05/3)

```

```

[1] 15

> con2_adj_p <- p.adjust(myresult_F$permutation_p[, 2], "BH")
> sum(con2_adj_p<=0.05/3)

[1] 15

> con3_adj_p <- p.adjust(myresult_F$permutation_p[, 3], "BH")
> sum(con3_adj_p<=0.05/3)

[1] 11

> which(con2_adj_p<=0.05/3)

[1] 64 92 98 132 476 580 636 653 657 802 831 881 898 932 992

> which(con3_adj_p<=0.05/3)

[1] 64 88 98 304 324 468 476 580 868 898 992

> unifdata_F <- matrix(runif(1000*18, 0.15, 0.98), 1000, 18)
> mydesign2_F <- c(rep(0, 6), rep(1, 6), rep(2, 6))
> aContrast <- c("X1-X0", "X2-X1", "X2-X0")
> myresult2_F <- RBM_F(unifdata_F, mydesign2_F, aContrast, 100, 0.05)
> summary(myresult2_F)

      Length Class  Mode
ordfit_t      3000  -none- numeric
ordfit_pvalue 3000  -none- numeric
ordfit_beta1  3000  -none- numeric
permutation_p 3000  -none- numeric
bootstrap_p    3000  -none- numeric

> sum(myresult2_F$bootstrap_p[, 1]<=0.05)

[1] 55

> sum(myresult2_F$bootstrap_p[, 2]<=0.05)

[1] 57

> sum(myresult2_F$bootstrap_p[, 3]<=0.05)

[1] 61

> which(myresult2_F$bootstrap_p[, 1]<=0.05)

```

```

[1] 15 18 27 34 36 73 90 122 127 166 174 186 199 222 232 244 258 260 299
[20] 312 354 375 382 384 400 455 459 477 511 515 523 551 552 561 572 608 616 621
[39] 632 647 649 660 666 700 709 751 761 818 867 889 891 904 962 964 980

> which(myresult2_F$bootstrap_p[, 2]<=0.05)

[1] 15 18 19 36 62 73 82 87 90 122 127 166 174 186 199 222 232 258 299
[20] 312 354 375 384 400 455 459 464 477 485 515 523 551 552 561 572 608 616 621
[39] 632 647 649 666 696 700 709 739 751 800 816 818 838 850 867 904 962 964 989

> which(myresult2_F$bootstrap_p[, 3]<=0.05)

[1] 15 18 27 33 34 36 62 73 82 87 90 122 127 166 174 186 199 222 244
[20] 258 260 262 299 312 354 375 400 455 459 464 477 485 506 511 515 523 548 549
[39] 551 552 561 572 608 616 621 622 632 647 649 666 700 708 709 751 756 816 818
[58] 867 904 962 980

> con21_adj_p <- p.adjust(myresult2_F$bootstrap_p[, 1], "BH")
> sum(con21_adj_p<=0.05/3)

[1] 11

> con22_adj_p <- p.adjust(myresult2_F$bootstrap_p[, 2], "BH")
> sum(con22_adj_p<=0.05/3)

[1] 13

> con23_adj_p <- p.adjust(myresult2_F$bootstrap_p[, 3], "BH")
> sum(con23_adj_p<=0.05/3)

[1] 9

```

4 Ovarian cancer methylation example using the RBM_T function

Two-group comparisons are the most common contrast in biological and biomedical field. The ovarian cancer methylation example is used to illustrate the application of RBM_T in identifying differentially methylated loci. The ovarian cancer methylation example is taken from the genome-wide DNA methylation profiling of United Kingdom Ovarian Cancer Population Study (UKOPS). This study used Illumina Infinium 27k Human DNA methylation Beadchip v1.2 to obtain DNA methylation profiles on over 27,000 CpGs in whole blood cells from 266 ovarian cancer women and 274 age-matched healthy controls. The data are downloaded from the NCBI GEO website with access number GSE19711. For illustration purpose, we chose the first 1000 loci in 8 randomly selected women with 4 ovarian cancer cases (pre-treatment) and 4 healthy controls. The following codes show the process of generating significant differential DNA methylation loci using the RBM_T function and presenting the results for further validation and investigations.

```
> system.file("data", package = "RBM")
```

```
[1] "/private/var/folders/r0/l4fjk6cj5xj0j3brt4bplpl40000gt/T/RtmpN1QDWY/Rinst204b4c9f21f7/RBM/d
```

```
> data(ovarian_cancer_methylation)
> summary(ovarian_cancer_methylation)
```

IlmnID	Beta	exmdata2[, 2]	exmdata3[, 2]
cg00000292: 1	Min. :0.01058	Min. :0.01187	Min. :0.009103
cg00002426: 1	1st Qu.:0.04111	1st Qu.:0.04407	1st Qu.:0.041543
cg00003994: 1	Median :0.08284	Median :0.09531	Median :0.087042
cg00005847: 1	Mean :0.27397	Mean :0.28872	Mean :0.283729
cg00006414: 1	3rd Qu.:0.52135	3rd Qu.:0.59032	3rd Qu.:0.558575
cg00007981: 1	Max. :0.97069	Max. :0.96937	Max. :0.970155
(Other) :994		NA's :4	

exmdata4[, 2]	exmdata5[, 2]	exmdata6[, 2]	exmdata7[, 2]
Min. :0.01019	Min. :0.01108	Min. :0.01937	Min. :0.01278
1st Qu.:0.04092	1st Qu.:0.04059	1st Qu.:0.05060	1st Qu.:0.04260
Median :0.09042	Median :0.08527	Median :0.09502	Median :0.09362
Mean :0.28508	Mean :0.28482	Mean :0.27348	Mean :0.27563
3rd Qu.:0.57502	3rd Qu.:0.57300	3rd Qu.:0.52099	3rd Qu.:0.52240
Max. :0.96658	Max. :0.97516	Max. :0.96681	Max. :0.95974
	NA's :1		

exmdata8[, 2]
Min. :0.01357
1st Qu.:0.04387
Median :0.09282
Mean :0.28679
3rd Qu.:0.57217
Max. :0.96268

```
> ovarian_cancer_data <- ovarian_cancer_methylation[, -1]
> label <- c(1, 1, 0, 0, 1, 1, 0, 0)
> diff_results <- RBM_T(aData=ovarian_cancer_data, vec_trt=label, repetition=100, alpha=0.05)
> summary(diff_results)
```

	Length	Class	Mode
ordfit_t	1000	-none-	numeric
ordfit_pvalue	1000	-none-	numeric
ordfit_beta0	1000	-none-	numeric
ordfit_beta1	1000	-none-	numeric
permutation_p	1000	-none-	numeric
bootstrap_p	1000	-none-	numeric

```
> sum(diff_results$ordfit_pvalue<=0.05)
```

```
[1] 45
```

```
> sum(diff_results$permutation_p<=0.05)
```

```
[1] 61
```

```
> sum(diff_results$bootstrap_p<=0.05)
```

```
[1] 71
```

```
> ordfit_adj_p <- p.adjust(diff_results$ordfit_pvalue, "BH")
```

```
> sum(ordfit_adj_p<=0.05)
```

```
[1] 0
```

```
> perm_adj_p <- p.adjust(diff_results$permutation_p, "BH")
```

```
> sum(perm_adj_p<=0.05)
```

```
[1] 4
```

```
> boot_adj_p <- p.adjust(diff_results$bootstrap_p, "BH")
```

```
> sum(boot_adj_p<=0.05)
```

```
[1] 10
```

```
> diff_list_perm <- which(perm_adj_p<=0.05)
```

```
> diff_list_boot <- which(boot_adj_p<=0.05)
```

```
> sig_results_perm <- cbind(ovarian_cancer_methylation[diff_list_perm, ], diff_results$ordfit_t[diff_list_perm, ])
```

```
> print(sig_results_perm)
```

	IlmnID	Beta	exmdata2[, 2]	exmdata3[, 2]	exmdata4[, 2]
5	cg00006414	0.07635468	0.07442468	0.15698040	0.08676092
103	cg00094319	0.73784280	0.73532960	0.75574900	0.73830220
346	cg00331237	0.05972383	NA	0.08204769	0.08345662
848	cg00826384	0.05721674	0.05612171	0.06644259	0.06358381
	exmdata5[, 2]	exmdata6[, 2]	exmdata7[, 2]	exmdata8[, 2]	
5	0.07982556	0.08111396	0.08271889	0.08045977	
103	0.67349260	0.73510200	0.75715920	0.78981220	
346	0.05372019	0.06241126	0.06955040	0.09140985	
848	0.05230160	0.06119713	0.06542751	0.06240686	
	diff_results\$ordfit_t[diff_list_perm]				
5	-1.389459				
103	-2.268711				
346	-3.767916				
848	-2.314412				
	diff_results\$permutation_p[diff_list_perm]				
5	0				
103	0				
346	0				
848	0				

```
> sig_results_boot <- cbind(ovarian_cancer_methylation[diff_list_boot, ], diff_results$ordfit_t[
> print(sig_results_boot)
```

	IlmnID	Beta	exmdata2[, 2]	exmdata3[, 2]	exmdata4[, 2]
146	cg00134539	0.61101320	0.53321780	0.45999340	0.46787420
189	cg00176210	0.28756520	0.39161870	0.44272520	0.44725330
259	cg00234961	0.04192170	0.04321576	0.05707140	0.05327565
280	cg00260778	0.64319890	0.60488960	0.56735060	0.53150910
285	cg00263760	0.09050395	0.10197760	0.14801710	0.12242400
632	cg00615377	0.11265030	0.16140570	0.19404450	0.17468600
743	cg00717862	0.07999436	0.07873347	0.06089359	0.06171374
911	cg00888479	0.07388961	0.07361080	0.10149800	0.09985076
928	cg00901493	0.03737166	0.03903724	0.04684618	0.04981432
979	cg00945507	0.13432250	0.23854600	0.34749760	0.28903340
	exmdata5[, 2]	exmdata6[, 2]	exmdata7[, 2]	exmdata8[, 2]	
146	0.67191510	0.63137380	0.47929610	0.45428300	
189	0.34106080	0.33765930	0.41252110	0.37024890	
259	0.04030003	0.03996053	0.05086962	0.05445672	
280	0.61920530	0.61925200	0.46753250	0.55632410	
285	0.11693600	0.10650430	0.12281160	0.12310430	
632	0.12573100	0.14483660	0.16338240	0.20130510	
743	0.07594936	0.09062161	0.06475791	0.07271878	
911	0.08633986	0.06765189	0.09070268	0.12417730	
928	0.04490690	0.04204062	0.05050039	0.05268215	
979	0.11848510	0.16653850	0.30718420	0.26624740	
	diff_results\$ordfit_t[diff_list_boot]				
146		5.394750			
189		-3.097987			
259		-4.052697			
280		4.170347			
285		-3.093997			
632		-3.661161			
743		3.444684			
911		-3.621731			
928		-2.716443			
979		-4.750997			
	diff_results\$bootstrap_p[diff_list_boot]				
146		0			
189		0			
259		0			
280		0			
285		0			
632		0			
743		0			
911		0			
928		0			

979

0