

# Package ‘AnnotationFuncs’

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**License** GPL-2

## BugReports

**Title** Annotation translation functions

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**Description** Functions for handling translating between different identifiers using the Biocore Data Team data-packages (e.g. org.Bt.eg.db).

**Version** 1.26.0

**biocViews** AnnotationData, Software

**URL** <http://www.iysik.com/index.php?page=annotation-functions>

**Date** 2010-11-29

**Depends** R (>= 2.7.0), AnnotationDbi

**Imports** DBI

**Suggests** org.Bt.eg.db, GO.db, org.Hs.eg.db, hom.Hs.inp.db

**Collate** 'annotation-funcs.R' 'homologe.R'

**NeedsCompilation** no

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AnnotationFuncs-package

*Annotation translation functions*

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**Description**

Package: AnnotationFuncs  
Type: Package  
Version: 1.3.0  
Date: 2011-06-10  
License: GPL-2  
LazyLoad: yes

**Details**

Functions for handling translations between different identifiers using the Biocore Data Team data-packages (e.g. `org.Bt.eg.db`). Primary functions are `translate` for translating and `getOrthologs` for efficient lookup of homologues using the Inparanoid databases. Other functions include functions for selecting Refseqs or Gene Ontologies (GO).

**Author(s)**

Stefan McKinnon Edwards <stefan.hoj-edwards@agrsci.dk>

**References**

<http://www.iysik.com/index.php?page=annotation-functions>

**See Also**

`translate`, `getOrthologs`

**Examples**

```
library(org.Bt.eg.db)
gene.symbols <- c('DRBP1', 'SERPINA1', 'FAKE', 'BLABLA')
# Find entrez identifiers of these genes.
eg <- translate(gene.symbols, org.Bt.egSYMBOL2EG)
# Note that not all symbols were translated.

# Go directly to Refseq identifiers.
refseq <- translate(gene.symbols, from=org.Bt.egSYMBOL2EG, to=org.Bt.egREFSEQ)
# Pick the proteins:
pickRefSeq(refseq, priorities=c('NP', 'XP'), reduce='all')
```

---

.dbEscapeString      *Private Escape string...*

---

### Description

Private Escape string

### Usage

```
.dbEscapeString(str, raise.error=TRUE)
```

### Arguments

str                      String to test  
raise.error              Logical, whether to raise an error or not.

### Details

Does not escape strings, but raises an error if any character expect normal letters and underscores are found in the string.

### Value

Invisible logical

---

.getTableNames              *Gets the table name from the INPARANOID style genus names.*

---

### Description

Gets the table name from the INPARANOID style genus names.

### Usage

```
.getTableNames(genus)
```

### Arguments

genus                      5 character INPARANOID genus name, such as "BOSTA", "HOMSA" or "MUSMU".

### Details

The INPARANOID style genus name is a 5 letter acronym of the species name. Quote INPARANOID (?hom.Hs.inpBOSTA):

*Names for these maps are done in the "INPARANOID style" which means that they are normally the 1st three letters of the genus followed by the 1st two letters of the species. For example: "Mus musculus" becomes "MUSMU", "Homo sapiens" becomes "HOMSA", "Monodelphis domestica" becomes "MONDO" etc. This means that for most of these organisms it will be possible to easily guess the abbreviations used. An exception may occur in the future if a new model organism has a very similar genus and species name to an existing one.*

**Value**

Table name for genus.

**Author(s)**

Stefan McKinnon Edwards <stefanm.edwards@agrsci.dk>

**References**

<http://www.bioconductor.org/packages/release/bioc/html/AnnotationDbi.html>

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*.pickRef*

*Secret function that does the magic for pickRefSeq.*

---

**Description**

Secret function that does the magic for pickRefSeq.

**Usage**

```
.pickRef(l, priorities, reduce=c("all", "first", "last"))
```

**Arguments**

<code>l</code>	List.
<code>priorities</code>	How to prioritize.
<code>reduce</code>	How to reduce.

**Details**

Do not use it, use [pickRefSeq](#)!

**Value**

List.

**Note**

Hey, you found a secret function! Keep it that way!

**Author(s)**

Stefan McKinnon Edwards <stefan.hoj-edwards@agrsci.dk>

**See Also**

[pickRefSeq](#)

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getEvidenceCodes      *Returns GO evidence codes.*

---

**Description**

Returns GO evidence codes.

**Value**

Matrix of two columns, first column with codes, second column with description of codes.

**Author(s)**

Stefan McKinnon Edwards <stefan.hoj-edwards@agrsci.dk>

**References**

?org.Bt.egGO

**See Also**

[pickGO](#)

**Examples**

```
getEvidenceCodes()
```

---

getOrthologs      *Performs quicker lookup for orthologs in homologue data packages...*

---

**Description**

Performs quicker lookup for orthologs in homologue data packages

**Usage**

```
getOrthologs(values, mapping, genus, threshold=1, pre.from, pre.to,
             post.from, post.to, ...)
```

**Arguments**

values	Vector, coerced to character vector, of values needed mapping by homology.
mapping	Homology mapping object, such as <code>hom.Hs.inpBOSTA</code> or <code>revmap(hom.Hs.inpBOSTA)</code> .
genus	Character vector. 5 character INPARANOID style genus name of the mapping object, e.g. 'BOSTA' for both <code>hom.Hs.inpBOSTA</code> and <code>revmap(hom.Hs.inpBOSTA)</code> .
threshold	Numeric value between 0 and 1. Only clustered homologues with a pairwise score above the threshold is included. The native implementation has this set to 1.

pre.from	Mapping object if values needs translation before mapping. E.g. values are entrez and hom.Hs.inpBOSTA requires ENSEMBLPROT, hom.Hs.inpAPIME requires Refseq (?). Arguments from and to are just like in <a href="#">translate</a> .
pre.to	Second part of translation before mapping.
post.from	Translate the result from homology mapping to a desired id; just like in <a href="#">translate</a> .
post.to	Second part of translation after mapping.
...	Additional arguments sent to <a href="#">translate</a> .

### Details

Using the INPARANOID data packages such as hom.Hs.inp.db is very, very slow and can take up to 11 min (on this particular developers workstation). This function introduces a new method that can do it in just 20 seconds (on the developers workstation). In addition, it includes options for translating between different identifiers both before and after the mapping.

### Value

List. Names of list corresponds to values, except those that could not be mapped nor translated. Entries are character vectors.

### Author(s)

Stefan McKinnon Edwards <stefan.hoj-edwards@agrsci.dk>

### References

?hom.Hs.inp.db - <http://inparanoid.sbc.su.se/>

Berglund, A.C., Sjolund, E., Ostlund, G., Sonnhammer, E.L.L. (2008) InParanoid 6: eukaryotic ortholog clusters with inparalogs *Nucleic Acids Res.* **36**:D263–266

O'Brien, K.P., Mairo, R., Sonnhammer, E.L.L (2005) Inparanoid: A Comprehensive Database of Eukaryotic Orthologs *NAR* **33**:D476–D480

Remm, M., Storm, C.E.V, Sonnhammer, E.L.L (2001) Automatic clustering of orthologs and inparalogs from pairwise species comparisons *J. Mol. Biol.* **314**:1041–1052

### See Also

[translate](#), [.getTableNames](#), [mapLists](#)

### Examples

```
library(hom.Hs.inp.db)
library(org.Hs.eg.db)
library(org.Bt.eg.db)
getOrthologs("ENSBTAP0000024572", revmap(hom.Hs.inpBOSTA), 'BOSTA')
# And now, we will map from entrez genes 1, 2 and 3 to bovine Refseq
bovine.ensembl <- getOrthologs(c(1,2,3), hom.Hs.inpBOSTA, 'BOSTA', pre.from=org.Hs.egENSEMBLPROT, post.from=
refseqs <- translate(unlist(bovine.ensembl, use.names=FALSE), org.Bt.egREFSEQ)
hs2bt.refseqs <- mapLists(bovine.ensembl, refseqs)
# Another way of doing it:
hs2bt.refseqs2 <- lapply(bovine.ensembl, translate, from=org.Bt.egREFSEQ, simplify=TRUE) # simplify=TRUE is
```

---

mapLists	<i>Replaces contents of list A with elements of list B...</i>
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---

## Description

Replaces contents of list A with elements of list B

## Usage

```
mapLists(A, B, removeNAs=TRUE)
```

## Arguments

A	List, elements are coerced to character for mapping to B.
B	List.
removeNAs	Boolean, whether to remove the NAs that occur because an element was not found in B.

## Details

Combines two lists, A and B, such that names(A) are preserved, mapping to the values of B, using names(B) as look up. Ie. replaces values in A with values in B, using names(B) as look up for values in A. Once more? See examples. *NB!* None-mapped entries are returned as NA, but can be removed using [removeNAs](#).

## Value

List.

## Author(s)

Stefan McKinnon Edwards <stefan.hoj-edwards@agrsci.dk>

## See Also

[removeNAs](#)

## Examples

```
A <- list('a1'='alpha', 'a2'='beta', 'a3'=c('gamma', 'delta'))
B <- list('alpha'='b1', 'gamma'=c('b2', 'b3'), 'delta'='b4')
mapLists(A, B)
```

---

pickGO *Cleans up result from org...*

---

### Description

Cleans up result from org.Xx.egGO and returns specific GO identifiers

### Usage

```
pickGO(l, evidence=NA, category=NA)
```

### Arguments

l	Character vector, or list of, og GO identifiers.
evidence	Character vector, filters on which kind of evidence to return; for a larger list see <a href="#">getEvidenceCodes</a> . \* Evidence codes may be: c('IMP', 'IGI', 'IPI', 'ISS', 'IDA', 'IEP', 'IEA') \* Leave as NA to ignore filtering on this part.
category	Character vector, filters on which ontology to return: biological process (BP), cellular component (CC), or molecular function (MF). \* Leave as NA to ignore filtering on this part.

### Details

Cleans up result from org.Xx.egGO and returns GO identifier for either biological process (BP), cellular component (CC), or molecular function (MF). Can be used on list of GOs from [translate](#), or a single list of GOs from an annotation package. May reduce list, if the (sub)list does not contain the chosen class!

### Value

List with only the picked elements.

### Author(s)

Stefan McKinnon Edwards <stefan.hoj-edwards@agrsci.dk>

### See Also

[pickRefSeq](#), [getEvidenceCodes](#), [translate](#)

### Examples

```
library(org.Bt.eg.db)
genes <- c(280705, 280706, 100327208)
GO <- translate(genes, org.Bt.egGO)
# Get all biological processes:
pickGO(GO, category='BP')
# Get all ontologies with experimental evidence:
pickGO(GO, evidence=c('IMP', 'IGI', 'IPI', 'ISS', 'IDA', 'IEP', 'IEA'))
```



---

pickRefSeq	<i>Picks a prioritised RefSeq identifier from a list of identifiers...</i>
------------	--

---

### Description

Picks a prioritised RefSeq identifier from a list of identifiers

### Usage

```
pickRefSeq(l, priorities=c("NP", "XP", "NM", "XM"), reduce=c("all",  
  "first", "last"))  
pickRefSeq.mRNA(l)  
pickRefSeq.Protein(l)
```

### Arguments

l	Vector or list of RefSeqs accessions to pick from. If list given, applies the prioritisation to each element in the list.
priorities	Character vector of prioritised prefixes to pick by. Eg. <code>c("NP", "NM")</code> returns RefSeqs starting 'NP', and if none found, those starting 'NM'. If no RefSeqs are found according to the priorities, Null is returned, unless the last element in priorities is '*'. Uses <code>grepl</code> , so see these for pattern matching. Default: <code>c('NP','XP','NM','XM')</code>
reduce	Reducing method, either return all annotations (one-to-many relation) or the first or last found annotation. The reducing step is applied after translating to the goal: <code>all</code> : returns all annotations <code>first</code> or <code>last</code> : choose first or last of arbitrarily ordered list.

### Details

When translating to RefSeq, typically multiple identifiers are returned, referring to different types of products, such as genomic molecule, mature mRNA or the protein, and they can be predicted, properties that can be read from the prefix (<http://www.ncbi.nlm.nih.gov/refseq/key.html>). E.g. "XM\_" is predicted mRNA and "NP\_" is a protein. Run `?org.Bt.egREFSEQ`.

### Value

If vector given, returns vector. If list given, returns list without element where nothing could be picked.

### Author(s)

Stefan McKinnon Edwards <[stefan.hoj-edwards@agrsci.dk](mailto:stefan.hoj-edwards@agrsci.dk)>

### Examples

```
library(org.Bt.eg.db)  
symbols <- c("SERPINA1", "KERA", "CD5")  
refseq <- translate(symbols, from=org.Bt.egSYMBOL2EG, to=org.Bt.egREFSEQ)  
mRNA <- pickRefSeq(refseq, priorities=c('NM','XM'))  
proteins <- pickRefSeq(refseq, priorities=c('NP','XP'))  
# The same.
```

```
mRNA <- pickRefSeq.mRNA(refseq)
proteins <- pickRefSeq.Protein(refseq)
```

---

removeNAs *Removes entries equal NA from list or vector...*

---

### Description

Removes entries equal NA from list or vector

### Usage

```
removeNAs(l)
```

### Arguments

l                    Vector or list.

### Details

Removes entries equal NA, but not mixed entries containing, amongst others, NA. Good for use after [mapLists](#) that might return entries equal NA.

### Author(s)

Stefan McKinnon Edwards <stefan.hoj-edwards@agrsci.dk>

### Examples

```
removeNAs(list('a'=NA, 'b'=c(NA, 'B'), 'c'='C'))
```

---

translate *Translate between different identifiers...*

---

### Description

Translate between different identifiers

### Usage

```
translate(values, from, to, reduce=c("all", "first", "last"),
          return.list=TRUE, remove.missing=TRUE, simplify=FALSE, ...)
```

**Arguments**

values	Vector of annotations that needs translation. Coerced to character vector.
from	Type of annotation values are given in. NB! take care in the orientation of the package, ie. if you have RefSeq annotations, use <code>org.Bt.egREFSEQ2EG</code> or (in some cases) <code>revmap(org.Bt.egREFSEQ)</code> .
to	Desired goal, eg. <code>org.Bt.egENSEMBLPROT</code> . If NULL (default), goal if the packages primary annotation (eg. <code>entrez gene</code> for <code>org.Bt.eg.db</code> ). Throws a warning if the organisms in <code>from</code> and <code>to</code> are not the same.
reduce	Reducing method, either return all annotations (one-to-many relation) or the first or last found annotation. The reducing step is applied after translating to the goal: <code>all</code> : returns all annotations <code>first</code> or <code>last</code> : choose first or last of arbitrarily ordered list.
return.list	Logical, when TRUE, returns the translation as a list where names
remove.missing	Logical, whether to remove non-translated values, defaults TRUE.
simplify	Logical, unlists the result. Defaults to FALSE. Usefull when using <code>translate</code> in a <code>lapply</code> or <code>sapply</code> .
...	Additional arguments sent to <code>pickGO</code> if <code>from</code> returns GO set.

**Details**

Function for translating from one annotation to another, eg. from RefSeq to Ensemble. This function takes a vector of annotation values and translates first to the primary annotation in the Biocore Data Team package (ie. `entrez gene` identifier for `org.Bt.eg.db`) and then to the desired product, while removing non-translated annotations and optionally reducing the result so there is only a one-to-one relation.

If you want to do some further mapping on the result, you will have to use either `unlist` or `lapply`, where the first returns all the end-products of the first mapping, returning a new list, and the latter produces a list-within-list.

If `from` returns GO identifiers (e.g. `from = org.Bt.egGO`), then the returned resultset is more complex and consists of several layers of lists instead of the usual list of character vectors. If `to` has also been specified, the GO IDs must be extracted (internally) and you have the option of filtering for evidence and category at this point. See `pickGO`.

**Value**

List; names of elements are values and the elements are the translated elements, or NULL if not translatable with `remove.missing = TRUE`.

**Note**

Requires user to deliver the annotation packages such as `org.Bt.egREFSEQ`.

**Author(s)**

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**See Also**

[pickRefSeq](#), [pickGO](#)

**Examples**

```
library(org.Bt.eg.db)
genes <- c(280705, 280706, 100327208)
translate(genes, org.Bt.egSYMBOL)

symbols <- c("SERPINA1", "KERA", "CD5")
refseq <- translate(symbols, from=org.Bt.egSYMBOL2EG, to=org.Bt.egREFSEQ)
# Pick the proteins:
pickRefSeq(refseq, priorities=c('NP', 'XP'), reduce='all')

# If you wanted do do some further mapping on the result from
# translate, simply use lapply.

library(GO.db)
GO <- translate(genes, org.Bt.egGO)
# Get all biological processes:
pickGO(GO, category='BP')
# Get all ontologies with experimental evidence:
pickGO(GO, evidence=c('IMP', 'IGI', 'IPI', 'ISS', 'IDA', 'IEP', 'IEA'))
```

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