Package 'prabclus'

September 24, 2024

Title Functions for Clustering and Testing of Presence-Absence, Abundance and Multilocus Genetic Data

Version 2.3-4

Date 2024-09-24

Depends R (>= 2.10), MASS, mclust

Suggests spdep, spatialreg, bootstrap, foreign, mvtnorm

Description Distance-based parametric bootstrap tests for clustering with spatial neighborhood information. Some distance measures, Clustering of presence-absence, abundance and multilocus genetic data for species delimitation, nearest neighbor based noise detection. Genetic distances between communities. Tests whether various distance-based regressions are equal. Try package?prabclus for on overview.

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NeedsCompilation no

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Repository CRAN

Date/Publication 2024-09-24 11:10:02 UTC

Contents

`prabclus-package` *prabclus package overview*

Description

Here is a list of the main functions in package prabclus. Most other functions are auxiliary functions for these.

Initialisation

prabinit Initialises presence/absence-, abundance- and multilocus data with dominant markers for use with most other key prabclus-functions.

alleleinit Initialises multilocus data with codominant markers for use with key prabclus-functions.

alleleconvert Generates the input format required by `alleleinit`.

Tests for clustering and nestedness

prabtest Computes the tests introduced in Hausdorf and Hennig (2003) and Hennig and Hausdorf (2004; these tests occur in some further publications of ours but this one is the most detailed statistical reference) for presence/absence data. Allows use of the geco-dissimilarity (Hennig and Hausdorf, 2006).

abundtest Computes the test introduced in Hausdorf and Hennig (2007) for abundance data.

homogen.test A classical distance-based test for homogeneity going back to Erdos and Renyi (1960) and Ling (1973).

Clustering

prabclust Species clustering for biotic element analysis (Hausdorf and Hennig, 2007, Hennig and Hausdorf, 2004 and others), clustering of individuals for species delimitation (Hausdorf and Hennig, 2010) based on Gaussian mixture model clustering with noise as implemented in R-package `mclust`, Fraley and Raftery (1998), on output of multidimensional scaling from distances as computed by `prabinit` or `alleleinit`. See also `stressvals` for help with choosing the number of MDS-dimensions.

hprabclust An unpublished alternative to `prabclust` using hierarchical clustering methods.

lociplots Visualisation of clusters of genetic markers vs. clusters of species.

NNclean Nearest neighbor based classification of observations as noise/outliers according to Byers and Raftery (1998).

Dissimilarity matrices

alleledist Shared allele distance (see the corresponding help pages for references).

dicedist Dice distance.

geco geco coefficient, taking geographical distance into account.

jaccard Jaccard distance.

kulczynski Kulczynski dissimilarity.

qkulczynski Quantitative Kulczynski dissimilarity for abundance data.

Communities

communities Constructs communities from geographical distances between individuals.

communitydist chord-, phiPT- and various versions of the shared allele distance between communities.

Tests for equality of dissimilarity-based regression

regeqdist Jackknife-based test for equality of two independent regressions between distances (Hausdorf and Hennig 2019).

regdistbetween Jackknife-based test for equality of regression involving all distances and regression involving within-group distances only (Hausdorf and Hennig 2019).

regdistbetweenone Jackknife-based test for equality of regression involving within-group distances of a reference group only and regression involving between-group distances (Hausdorf and Hennig 2019).

Small conversion functions

coord2dist Computes geographical distances from geographical coordinates.

geo2neighbor Computes a neighborhood list from geographical distances.

alleleconvert A somewhat restricted function for conversion of different file formats used for genetic data with codominant markers.

Data sets

kykladspecreg, siskiyou, veronica, tetragonula.

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References

Byers, S. and Raftery, A. E. (1998) Nearest-Neighbor Clutter Removal for Estimating Features in Spatial Point Processes, *Journal of the American Statistical Association*, 93, 577-584.

Erdos, P. and Renyi, A. (1960) On the evolution of random graphs. *Publications of the Mathematical Institute of the Hungarian Academy of Sciences* 5, 17-61.

Fraley, C. and Raftery, A. E. (1998) How many clusters? Which clusterin method? - Answers via Model-Based Cluster Analysis. *Computer Journal* 41, 578-588.

Hausdorf, B. and Hennig, C. (2003) Nestedness of north-west European land snail ranges as a consequence of differential immigration from Pleistocene glacial refuges. *Oecologia* 135, 102-109.

Hausdorf, B. and Hennig, C. (2007) Null model tests of clustering of species, negative co-occurrence patterns and nestedness in meta-communities. *Oikos* 116, 818-828.

Hausdorf, B. and Hennig, C. (2010) Species Delimitation Using Dominant and Codominant Multilocus Markers. *Systematic Biology*, 59, 491-503.

Hausdorf, B. and Hennig, C. (2019) Species delimitation and geography. Submitted.

Hennig, C. and Hausdorf, B. (2004) Distance-based parametric bootstrap tests for clustering of species ranges. *Computational Statistics and Data Analysis* 45, 875-896.

Hennig, C. and Hausdorf, B. (2006) A robust distance coefficient between distribution areas incorporating geographic distances. *Systematic Biology* 55, 170-175.

Ling, R. F. (1973) A probability theory of cluster analysis. *Journal of the American Statistical Association* 68, 159-164.

`abundtest` *Parametric bootstrap test for clustering in abundance matrices*

Description

Parametric bootstrap test of a null model of i.i.d., but spatially autocorrelated species against clustering of the species' population patterns. Note that most relevant functionality of `prabtest` (except of the use of the geco distance) is also included in `abundtest`, so that `abundtest` can also be used on binary presence-absence data. In spite of the lots of parameters, a standard execution (for the default test statistics, see parameter `teststat` below) will be

```
prabmatrix <- prabinit(file="path/abundmatrixfile", neighborhood="path/neighborhoodfile")
test <- abundtest(prabmatrix)
summary(test)
```

Note: Data formats are described on the `prabinit` help page. You may also consider the example datasets `kykladspecreg.dat` and `nb.dat`. Take care of the parameter `rows.are.species` of `prabinit`.

Usage

```
abundtest(prabobj, teststat = "distratio", tuning = 0.25,
                       times = 1000, p.nb = NULL,
                       prange = c(0, 1), nperp = 4, step = 0.1, step2 = 0.01,
```

```
                    twostep = TRUE, species.fixed=TRUE, prab01=NULL,
                    groupvector=NULL,
                    sarestimate=prab.sarestimate(prabobj),
                    dist = prabobj$distance,
                    n.species = prabobj$n.species)
```

Arguments

prabobj	an object of class prab (presence-absence data), as generated by prabinit.
teststat	string, indicating the test statistics. "isovertice": number of isolated vertices in the graph of tuning smallest distances between species. "lcomponent": size of largest connectivity component in this graph. "distratio": ratio between tuning smallest and largest distances. "nn": average distance of species to tuningth nearest neighbor. "inclusions": number of inclusions between areas of different species (tests for nestedness structure, not for clustering, and treats abundance matrices as presence-absence-data). "mean": mean of the distances between species (this is a rough measure of species co-occurrence). "groups": this requires a specification of a vector defining different groups of species via parameter groupvector. The test statistic is then the mean of the distances between species of the same group. This is computed over all species, but also for every single group of species. It also includes the "mean"-test, so that the number of tests carried out is number of species groups with more than one element plus two.
tuning	integer or (if teststat="distratio") numerical between 0 and 1. Tuning constant for test statistics, see teststat.
times	integer. Number of simulation runs.
p.nb	numerical between 0 and 1. The probability that a new region is drawn from the non-neighborhood of the previous regions belonging to a species under generation. If NULL (the default), and prabobj$spatial, prabtest estimates this by function autoconst. Otherwise the next five parameters have no effect. If NULL, and !prabobj$spatial, spatial structure is ignored.
prange	numerical range vector, lower value not smaller than 0, larger value not larger than 1. Range where pd is to be found. Used by function autoconst.
nperp	integer. Number of simulations per pd-value. Used by function autoconst.
step	numerical between 0 and 1. Interval length between subsequent choices of pd for the first simulation. Used by function autoconst.
step2	numerical between 0 and 1. Interval length between subsequent choices of pd for the second simulation (see parameter twostep). Used by function autoconst.
twostep	logical. If TRUE, a first estimation step for pd is carried out in the whole prange, and then the final estimation is determined between the preliminary estimator -5*step2 and +5*step2. Else, the first simulation determines the final estimator. Used by function autoconst.
species.fixed	logical. Indicates if the range sizes of the species are held fixed in the test simulation (TRUE) or generated from their empirical distribution in x (FALSE) for presence-absence data. See function randpop.nb. Use always TRUE for abundance data (not necessary if teststat="inclusions").

prab01	prabinit-object based on presence-absence matrix of same dimensions than the abundance matrix of prabobj. This specifies the presences and absences on which the presence/absence step of abundance-based tests is based (see details). If NULL (which is usually the only reasonable choice), prab01 is computed in order to indicate the nonzeroes of prabobj$prab.
groupvector	integer vector. For every species, a number indicating the species' group membership. Needed only if teststat="groups".
sarestimate	Estimator of the parameters of a simultaneous autoregression model corresponding to the null model for abundance data from Hausdorf and Hennig (2007) as generated by prab.sarestimate. This requires package spdep. Note that by explicitly specifying sarestimate=NULL simulation of 0-1 matrices can be enforced.
dist	One of "jaccard", "kulczynski", "qkulczynski" or "logkulczynski" specifying the distance measure on which the test is based. By default, this is taken from prabobj.
n.species	number of species. By default this is taken from prabobj. This should normally not be changed.

Details

For presence-absence data, the routine is described in prabtest. For abundance data, the first step under the null model is to simulated presence-absence patterns as in prabtest. The second step is to fit a simultaneous autoregression (SAR) model (Ripley 1981, section 5.2) to the log-abundances, see prab.sarestimate. The simulation from the null model is implemented in regpop.sar. For more details see Hennig and Hausdorf (2004) for presence-absence data and Hausdorf and Hennig (2007) for abundance data and the test statistics "mean" and "groups", which can also be applied to binary data.

If p.nb=NA was specified, a diagnostic plot for the estimation of pd is plotted by autoconst. For details see Hennig and Hausdorf (2004) and the help pages of the cited functions.

Value

An object of class prabtest, which is a list with components

results	vector of test statistic values for all simulated populations. For teststat="groups" a list with components overall (means of within group-distances), mean (means of all distances), gr (matrix with a row for every group, giving the groupwise within-group distance means).
p.above	p-value against an alternative that generates large values of the test statistic (usually reasonable for teststat="inclusions", "groups", "mean").
p.below	p-value against an alternative that generates small values of the test statistic (usually reasonable for "lcomponent", "nn", "distratio"; for "isovertice", the two-sided p may make sense which is twice the smaller one of p.above and p.below).
datac	test statistic value for the original data. (specgroups-output for teststat="groups").
tuning	see above.

distance	dist above.
teststat	see above.
pd	p.nb above.
abund	TRUE if simultaneous autoregression has been used (i.e., a sarestimate has been supplied or computed).
sarlambda	Estimator of the autocorrelation parameter lambda (see errorsarlm) defined so that the average weight of neighbors (see nb2listw) is standardized to 1.
sarestimate	the output object of prab.sarestimate.
groupinfo	list containing information from "groups" tests, with components lg (levels of groupvector), ng (number of groups), nsg (vector of group sizes), testm (value of "means" test statistic for input prabobj), pa (group-wise p.above), pb (group-wise p.below), pma (p.above of "means" test), pmb (p.below of "means" test).

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References

Hausdorf, B. and Hennig, C. (2007) Null model tests of clustering of species, negative co-occurrence patterns and nestedness in meta-communities. *Oikos* 116, 818-828.

Hennig, C. and Hausdorf, B. (2004) Distance-based parametric bootstrap tests for clustering of species ranges. *Computational Statistics and Data Analysis* 45, 875-896. http://stat.ethz.ch/Research-Reports/110.html.

Ripley, B. D. (1981) *Spatial Statistics*. Wiley.

See Also

prabinit generates objects of class prab.

autoconst estimates pd from such objects.

prabtest (analogous function for presence-absence data).

regpop.sar generates populations from the null model.

prab.sarestimate (parameter estimators for simultaneous autoregression model). This calls

errorsarlm (original estimation function from package spdep).

Some more information on the test statistics is given in homogen.test, lcomponent, distratio, nn, incmatrix.

Summary and print methods: summary.prabtest.

Examples

```
# Note: NOT RUN.
# This needs package spdep and a bunch of packages that are
# called by spdep!
# data(siskiyou)
# set.seed(1234)
# x <- prabinit(prabmatrix=siskiyou, neighborhood=siskiyou.nb,
#              distance="logkulczynski")
# a1 <- abundtest(x, times=5, p.nb=0.0465)
# a2 <- abundtest(x, times=5, p.nb=0.0465, teststat="groups",
#                groupvector=siskiyou.groups)
# These settings are chosen to make the example execution
# faster; usually you will use abundtest(x).
# summary(a1)
# summary(a2)
```

`allele2zeroone` *Converts alleleobject into binary matrix*

Description

Converts `alleleobject` with codominant markers into binary matrix with a column for each marker.

Usage

```
allele2zeroone(alleleobject)
```

Arguments

`alleleobject`	object of class `alleleobject` as generated by `alleleinit`.

Value

A 0-1-matrix with individuals as rows and markers (alleles) as columns.

Author(s)

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Examples

```
data(tetragonula)
ta <- alleleconvert(strmatrix=tetragonula[21:50,])
tai <- alleleinit(allelematrix=ta)
allele2zeroone(tai)
```

`alleleconvert` *Format conversion for codominant marker data*

Description

Codominant marker data (which here means: data with several diploid loci; two alleles per locus) can be represented in various ways. This function converts the formats `"genepop"` and `"structure"` into `"structurama"` and `"prabclus"`. `"genepop"` is a version of the format used by the package GENEPOP (Rousset, 2008), `"structure"` is a version of what is used by STRUCTURE (Pritchard et al., 2000), another one is `"structureb"`. `"structurama"` is a version of what is used by STRUCTURAMA (Huelsenbeck and Andolfatto, 2007) and `"prabclus"` is required by the function `alleleinit` in the present package.

Usage

```
alleleconvert(file=NULL,strmatrix=NULL, format.in="genepop",
                        format.out="prabclus",
                        alength=3,orig.nachar="000",new.nachar="-",
                        rows.are.individuals=TRUE, firstcolname=FALSE,
                        aletters=intToUtf8(c(65:90,97:122),multiple=TRUE),
                        outfile=NULL,skip=0)
```

Arguments

`file`	string. Filename of input file, see details. One of `file` and `strmatrix` needs to be specified.
`strmatrix`	matrix or data frame of strings, see details. One of `file` and `strmatrix` needs to be specified.
`format.in`	string. One of `"genepop"`, `"structure"`, or `"structureb"`, see details.
`format.out`	string. One of `"structurama"` or `"prabclus"`, see details.
`alength`	integer. If `format.in="genepop"`, length of code for a single allele.
`orig.nachar`	string. Code for missing values in input data.
`new.nachar`	string. Code for missing values in output data.
`rows.are.individuals`	logical. If `TRUE`, rows are interpreted as individuals and columns (variables if `strmatrix` is a data frame) as loci.
`firstcolname`	logical. If `TRUE`, it is assumed that the first column contains row names.
`aletters`	character vector. String of default characters for alleles if `format.out=="prabclus"` (the default is fine unless there is a locus that can have more than 62 different alleles in the dataset).
`outfile`	string. If specified, the output matrix (omitting quotes) is written to a file of this name (including row names if `fistcolname==TRUE`).
`skip`	number of rows to be skipped when reading data from a file (`skip`-argument of `read.table`).

Details

The formats are as follows (described is the format within R, i.e., for the input, the format of `strmatrix`; if `file` is specified, the file is read with `read.table(file,colClasses="character")` and should give the format explained below - note that `colClasses="character"` implies that quotes are not needed in the input file):

genepop Alleles are coded by strings of length `alength` and there is no space between the two alleles in a locus, so a value of `"258260"` means that in the corresponding locus the two alleles have codes 258 and 260.

structure Alleles are coded by strings of arbitrary length. Two rows correspond to each inidividual, the first row containing the first alleles in all loci and the second row containing the second ones.

structureb Alleles are coded by strings of arbitrary length. One row corresponds to each inidividual, containing first and second alleles in all loci (first and second allele of first locus, first and second allele of second locus etc.). This starts in the third row (first two have locus names and other information).

structurama Alleles are coded by strings of arbitrary length. the two alleles in each locus are written with brackets around them and a comma in between, so `"258260"` in `"genepop"` corresponds to `"(258,260)"` in `"structurama"`.

prabclus Alleles are coded by a single character and there is no space between the two alleles in a locus (e.g., `"AC"`).

Value

A matrix of strings in the format specified as `format.out` with an attribute `"alevels"`, a vector of all used allele codes if `format.out=="prabclus"`, otherwise vector of allele codes of last locus.

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References

Huelsenbeck, J. P., and P. Andolfatto (2007) Inference of population structure under a Dirichlet process model. *Genetics* 175, 1787-1802.

Pritchard, J. K., M. Stephens, and P. Donnelly (2000) Inference of population structure using multilocus genotype data. *Genetics* 155, 945-959.

Rousset, F. (2008) genepop'007: a complete re-implementation of the genepop software for Windows and Linux. *Molecular Ecology Resources* 8, 103-106.

See Also

`alleleinit`

Examples

```
  data(tetragonula)
# This uses example data file Heterotrigona_indoFO.dat
  str(alleleconvert(strmatrix=tetragonula))
  strucmatrix <-
    cbind(c("I1","I1","I2","I2","I3","I3"),
    c("122","144","122","122","144","144"),c("0","0","21","33","35","44"))
  alleleconvert(strmatrix=strucmatrix,format.in="structure",
    format.out="prabclus",orig.nachar="0",firstcolname=TRUE)
  alleleconvert(strmatrix=strucmatrix,format.in="structure",
    format.out="structurama",orig.nachar="0",new.nachar="-9",firstcolname=TRUE)
```

---

alleledist *Shared allele distance for diploid loci*

Description

Shared allele distance for codominant markers (Bowcock et al., 1994). One minus proportion of alleles shared by two individuals averaged over loci (loci with missing values for at least one individual are ignored).

Usage

```
alleledist(allelelist,ni,np,count=FALSE)
```

Arguments

allelelist	a list of lists. In the "outer" list, there are np lists, one for each locus. In the "inner" list, for every individual there is a vector of two codes (typically characters, see alleleinit) for the two alleles in that locus. Such a list can be constructed by unbuild.charmatrix out of the charmatrix component of an output object of alleleinit.
ni	integer. Number of individuals.
np	integer. Number of loci.
count	logical. If TRUE, the number of the individual to be processed is printed.

Value

A symmetrical matrix of shared allele distances between individuals.

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References

Bowcock, A. M., Ruiz-Linares, A., Tomfohrde, J., Minch, E., Kidd, J. R., Cavalli-Sforza, L. L. (1994) High resolution of human evolutionary trees with polymorphic microsatellites. *Nature* 368, 455-457.

See Also

alleleinit, unbuild.charmatrix

Examples

```
data(tetragonula)
tnb <-
coord2dist(coordmatrix=tetragonula.coord[1:50,],cut=50,file.format="decimal2",neighbors=TRUE)
ta <- alleleconvert(strmatrix=tetragonula[1:50,])
tai <- alleleinit(allelematrix=ta,neighborhood=tnb$nblist,distance="none")
str(alleledist((unbuild.charmatrix(tai$charmatrix,50,13)),50,13))
```

---

alleleinit *Diploid loci matrix initialization*

Description

`alleleinit` converts genetic data with diploid loci as generated by `alleleconvert` into an object of class `alleleobject`. `print.alleleobject` is a print method for such objects.

Usage

```
alleleinit(file = NULL, allelematrix=NULL,
                        rows.are.individuals = TRUE,
    neighborhood = "none", distance = "alleledist", namode="variables",
                       nachar="-", distcount=FALSE)


## S3 method for class 'alleleobject'
print(x, ...)
```

Arguments

`file`	string. File name. File must be in `"prabclus"` format, see details. Either `file` or `allelematrix` needs to be specified.
`allelematrix`	matrix in `"prabclus"`-format as generated by `alleleconvert`, see details. Either `file` or `allelematrix` needs to be specified.
`rows.are.individuals`	logical. If `TRUE`, rows are interpreted as individuals and columns are interpreted as loci.

neighborhood	A string or a list with a component for every individual. The components are vectors of integers indicating neighboring individuals. An individual without neighbors should be assigned a vector numeric(0). If neighborhood is a filename, it is attempted to read such a list from a file, where every row should correspond to one region (such as example dataset nb.dat). If neighborhood="none", all neighborhoods are set to numeric(0). The neighborhood can be tested by nbtest for consistency.
distance	"alleledist" or "none". The distance measure between individuals to compute by alleleinit.
namode	one of "single", "individuals", "variables" , or "none". Determines whether a single probability for the entry to be missing is computed for a single locus of an individual ("single"), a vector of individual-wise probabilities for loci to be missing ("individuals"), a vector of loci-wise probabilities for individuals to be missing ("variables") or no missingness probability at all.
nachar	character denoting missing values.
distcount	logical. If TRUE, during distance computation individuals are counted on the screen.
x	object of class alleleobject.
...	necessary for print method.

Details

The required input format is the output format "prabclus" of alleleconvert. Alleles are coded by a single character, so diploid loci need to be pairs of characters without space between the two alleles (e.g., "AC"). The input needs to be an individuals*loci matrix or data frame (or a file that produces such a data frame by read.table(file,stringsAsFactors=FALSE))

Value

alleleinit produces an object of class alleleobject (note that this is similar to class prab; for example both can be used with prabclust), which is a list with components

distmat	distance matrix between individuals.
amatrix	data frame of input data with string variables in the input format, see details. Note that in the output for an individual the whole locus is declared missing if at least one of its alleles is missing in the input.
charmatrix	matrix of characters in which there are two rows for every individual corresponding to the two alleles in every locus (column). Entries are allele codes but missing values are coded as NA.
nb	neighborhood list, see above.
ext.nblist	a neighborhood list in which for every row in charmatrix the second row number corresponding to the neighboring individuals is listed.
n.variables	number of loci.
n.individuals	number of individuals.
n.levels	maximum number of different alleles in a locus.

n.species	identical to n.individuals used for compatibility with prabclust.
alevels	character vector with all used allele codes not including missing values.
leveldist	matrix in which rows are loci, columns are alleles and entries are frequencies of alleles per locus.
prab	useless matrix of number of factor levels corresponding to amatrix added for compatibility with objects of class prab.
regperspec	vector of row-wise sums of prab added for compatibility with objects of class prab.
specperreg	vector of column-wise sums of prab added for compatibility with objects of class prab.
distance	string denoting the chosen distance measure, see above.
namode	see above.
naprob	probability of missing values, numeric or vector, see documentation of argument namode.
nasum	number of missing entries (individual/loci) in amatrix.
nachar	see above.
spatial	logical. TRUE if a neighborhood was submitted.

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See Also

alleleconvert, alleledist, prabinit.

Examples

```
# Only 50 observations are used in order to have a fast example.
  data(tetragonula)
  tnb <-
 coord2dist(coordmatrix=tetragonula.coord[1:50,],cut=50,file.format="decimal2",neighbors=TRUE)
  ta <- alleleconvert(strmatrix=tetragonula[1:50,])
  tai <- alleleinit(allelematrix=ta,neighborhood=tnb$nblist)
  print(tai)
```

`allelepaircomp` *Internal: compares two pairs of alleles*

Description

Used for computation of the genetic distances `alleledist`.

Usage

```
allelepaircomp(allelepair1,allelepair2,method="sum")
```

Arguments

`allelepair1`	vector of two allele codes (usually characters), or `NA`.
`allelepair2`	vector of two allele codes (usually characters), or `NA`.
`method`	one of `"sum"` or `"geometrical"`.

Value

If `method=="sum"`, number of shared alleles (0, 1 or 2), or `NA`. If `method=="geometrical"`, 0, 0.5, `sqrt(0.5)` (in case that one of the allelepairs is double such as in `c("A","B"),c("A","A")`) or 1, or `NA`.

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See Also

`alleledist`

Examples

```
allelepaircomp(c("A","B"),c("A","C"))
```

autoconst *Spatial autocorrelation parameter estimation*

Description

Monte Carlo estimation of the disjunction/spatial autocorrelation parameter pd for the simulation model used in randpop.nb, used for tests for clustering of presence-absence data.

autoconst is the main function; autoreg performs the simulation and is executed within autoconst.

Usage

```
autoconst(x, prange = c(0, 1), twostep = TRUE, step1 = 0.1,
step2 = 0.01, plot = TRUE, nperp = 4, ejprob = NULL,
species.fixed = TRUE, pdfnb=FALSE, ignore.richness=FALSE)

autoreg(x, probs, ejprob, plot = TRUE, nperp = 4, species.fixed = TRUE,
pdfnb=FALSE, ignore.richness=FALSE)
```

Arguments

x	object of class prab as generated by prabinit. Presence-absence data to be analyzed.
prange	numerical range vector, lower value not smaller than 0, larger value not larger than 1. Range where the parameter is to be found.
twostep	logical. If TRUE, a first estimation step is carried out in the whole prange, and then the final estimation is determined between the preliminary estimator -5*step2 and +5*step2. Else, the first simulation determines the final estimator.
step1	numerical between 0 and 1. Interval length between subsequent choices of pd for the first simulation.
step2	numerical between 0 and 1. Interval length between subsequent choices of pd for the second simulation in case of twostep=TRUE.
plot	logical. If TRUE, a scatterplot of pd-values against resulting ejprob values (see below), with regression line and data value of ejprob is shown.
nperp	integer. Number of simulations per pd-value.
ejprob	numerical between 0 and 1. Observed disjunction probability for data x; if not specified in advance, it will be computed by autoconst.
species.fixed	logical. If TRUE, sizes of generated species match the species sizes in x, else they are generated from the empirical distribution of species sizes in x.
probs	vector of numericals between 0 and 1. pd values for the simulation.
pdfnb	logical. If TRUE, the probabilities of the regions are modified according to the number of neighboring regions in randpop.nb, see Hennig and Hausdorf (2002), p. 5.

ignore.richness
: logical. If TRUE, there is no assumption of species richnesses to differ between regions in the null model. Regionwise probabilities don't differ in the generation of null data.

Details

The spatial autocorrelation parameter pd of the model for the generation of presence-absence data sets used by randpop.nb can be estimated by use of the observed disjuction probability ejprob which is the sum of all species' connectivity components minus the number of species divided by the number of "presence" entries minus the number of species. This is done by a simulation of artificial data sets with characteristics of x and different pd-values, governed by prange, step1, step2 and nperp. ejprob is then calculated for all simulated populations. A linear regression of ejprob on pd is performed and the estimator of pd is determined by computing the inverse of the regression function for the ejprob-value of x.

Value

autoconst produces the same list as autoreg with additional component ejprob. The components are

pd	(eventually) estimated parameter pd.
coef	(eventually) estimated regression coefficients.
ejprob	see above.

Author(s)

Christian Hennig <christian.hennig@unibo.it> https://www.unibo.it/sitoweb/christian.hennig/en

References

Hausdorf, B. and Hennig, C. (2003) Biotic Element Analysis in Biogeography. To appear in *Systematic Biology*.

Hausdorf, B. and Hennig, C. (2003) Nestedness of north-west European land snail ranges as a consequence of differential immigration from Pleistocene glacial refuges. *Oecologia* 135, 102-109.

Hennig, C. and Hausdorf, B. (2004) Distance-based parametric bootstrap tests for clustering of species ranges. *Computational Statistics and Data Analysis* 45, 875-896.

See Also

randpop.nb, prabinit, con.comp

Examples

```
options(digits=4)
data(kykladspecreg)
data(nb)
set.seed(1234)
```

```
x <- prabinit(prabmatrix=kykladspecreg, neighborhood=nb)
ax <- autoconst(x,nperp=2,step1=0.3,twostep=FALSE)
```

`build.charmatrix` *Internal: create character matrix out of allele list*

Description

For use in `alleleinit`. Creates a matrix of characters in which there are two rows for every individual corresponding to the two alleles in every locus (column) out of a list of lists, such as required by `alleledist`.

Usage

```
build.charmatrix(allelelist,n.individuals,n.variables)
```

Arguments

`allelelist`	A list of lists. In the "outer" list, there are `n.variables` lists, one for each locus. In the "inner" list, for every individual there is a vector of two codes (typically characters, see `alleleinit`) for the two alleles in that locus.
`n.individuals`	integer. Number of individuals.
`n.variables`	integer. Number of loci.

Value

A matrix of characters in which there are two rows for every individual corresponding to the two alleles in every locus (column).

Author(s)

Christian Hennig <christian.hennig@unibo.it> https://www.unibo.it/sitoweb/christian.hennig/en

See Also

`alleleinit`, `unbuild.charmatrix`

Examples

```
alist <- list()
alist[[1]] <- list(c("A","A"),c("B","A"),c(NA,NA))
alist[[2]] <- list(c("A","C"),c("B","B"),c("A","D"))
build.charmatrix(alist,3,2)
```

`build.ext.nblist` *Internal: generates neighborhood list for diploid loci*

Description

This is for use in `alleleinit`. Given a neighborhood list of individuals, a new neighborhood list is generated in which there are two entries for each individual (entry 1 and 2 refer to individual one, 3 and 4 to individual 2 and so on). Neighborhoods are preserved and additionally the two entries belonging to the same individual are marked as neighbors.

Usage

```
build.ext.nblist(neighbors,n.individuals=length(neighbors))
```

Arguments

`neighbors`	list of integer vectors, where each vector contains the neighbors of an individual.
`n.individuals`	integer. Number of individuals.

Value

list with `2*n.inidividuals` vectors of integers as described above.

Author(s)

Christian Hennig <christian.hennig@unibo.it> https://www.unibo.it/sitoweb/christian.hennig/en

See Also

`alleleinit`

Examples

```
data(veronica)
vnb <- coord2dist(coordmatrix=veronica.coord[1:20,], cut=20,
  file.format="decimal2",neighbors=TRUE)
build.ext.nblist(vnb$nblist)
```

build.nblist *Generate spatial weights from prabclus neighborhood list*

Description

This generates a listw-object as needed for estimation of a simultaneous autoregression model in package spdep from a neighborhood list of the type generated in prabinit.

Usage

```
build.nblist(prabobj,prab01=NULL,style="C")
```

Arguments

prabobj	object of class prab.
prab01	presence-absence matrix of same dimensions than the abundance matrix of prabobj. This specifies the presences and absences on which the presence/absence step of abundance-based tests is based (see details). If NULL (which is usually the only reasonable choice), prab01 is computed in order to indicate the nonzeroes of prabobj$prab.
style	can take values "W", "B", "C", "U", and "S" though tests suggest that "C" should be chosen. See nb2listw.

Value

A 'listw' object with the following members:

style	see above.
neighbours	the neighbours list in spdep-format.
weights	the weights for the neighbours and chosen style, with attributes set to report the type of relationships (binary or general, if general the form of the glist argument), and style as above.

Author(s)

Christian Hennig <christian.hennig@unibo.it> https://www.unibo.it/sitoweb/christian.hennig/en

See Also

nb2listw (which is called)

Examples

```
# Not run; requires package spdep
# data(siskiyou)
# x <- prabinit(prabmatrix=siskiyou, neighborhood=siskiyou.nb,
#             distance="logkulczynski")
# build.nblist(x)
```

---

cluspop.nb *Simulation of presence-absence matrices (clustered)*

Description

Generates a simulated matrix where the rows are interpreted as regions and the columns as species, 1 means that a species is present in the region and 0 means that the species is absent. Species are generated in order to produce 2 clusters of species with similar ranges. Spatial autocorrelation of a species' presences is governed by the parameter p.nb and a list of neighbors for each region.

Usage

```
cluspop.nb(neighbors, p.nb = 0.5, n.species, clus.specs, reg.group,
grouppf = 10, n.regions = length(neighbors),
vector.species = rep(1, n.species), pdf.regions = rep(1/n.regions,
n.regions), count = TRUE, pdfnb = FALSE)
```

Arguments

neighbors	A list with a component for every region. The components are vectors of integers indicating neighboring regions. A region without neighbors (e.g., an island) should be assigned a list numeric(0).
p.nb	numerical between 0 and 1. The probability that a new region is drawn from the non-neighborhood of the previous regions belonging to a species under generation. Note that for a given presence-absence matrix, this parameter can be estimated by autoconst (called pd there).
n.species	integer. Number of species.
clus.specs	integer not larger than n.species. Number of species restricted to one of the two groups of regions defined by reg.group (called "clustered species" because this leads to more similar species ranges).
reg.group	vector of pairwise distinct integers not larger than n. regions. Defines a group of regions to which a part of the clus.specs clustered species is restricted (more or less, see grouppf). The other clustered species are restricted to the complementary regions.
grouppf	numerical. The probability of the region of a clustered species to belong to the corresponding group of regions is up-weighted by factor grouppf compared to the generation of "non-clustered" species.
n.regions	integer. Number of regions.

vector.species	vector of integers. The sizes (i.e., numbers of regions) of the species are generated randomly from the empirical distribution of vector.species.
pdf.regions	numerical vector of length n.species. The entries must sum up to 1 and give probabilities for the regions to be drawn during the generation of a species. These probabilities are used conditional on the new region being a neighbor or a non-neighbor of the previous regions of the species, see p.nb, modified by grouppf for the clustered species.
count	logical. If TRUE, the number of the currently generated species is printed.
pdfnb	logical. If TRUE, the probabilities of the regions are modified according to the number of neighboring regions by dividing them relative to the others by min(1,number of neighbors).

Details

The non-clustered species are generated as explained on the help page for randpop.nb. The general principle for the clustered species is the same, but with modified probabilities for the regions. For each clustered species, one of the two groups of regions is drawn, distributed according to the sum of its regions' probability given by pdf.regions. The first region of such a species is only drawn from the regions of this group.

Value

A 0-1-matrix, rows are regions, columns are species.

Author(s)

Christian Hennig <christian.hennig@unibo.it> https://www.unibo.it/sitoweb/christian.hennig/en

References

Hennig, C. and Hausdorf, B. (2004) Distance-based parametric bootstrap tests for clustering of species ranges. *Computational Statistics and Data Analysis* 45, 875-896.

See Also

randpop.nb,

autoconst estimates p.nb from matrices of class prab. These are generated by prabinit.

Examples

```
data(nb)
set.seed(888)
cluspop.nb(nb, p.nb=0.1, n.species=10, clus.specs=9, reg.group=1:17,
vector.species=c(10))
```

communities *Construct communities from individuals*

Description

Construct communities from individuals using geographical distance and hierarchical clustering. Communities are clusters of geographically close individuals, formed by hclust with specified distance cutoff.

Usage

```
communities(geodist,grouping=NULL,
                        cutoff=1e-5,method="single")
```

Arguments

geodist	dist-object or matrix of geographical distances between individuals.
grouping	something that can be coerced into a factor. Different groups indicated by grouping cannot be together in the same community. (If NULL, there is no constraint.)
cutoff	numeric; clustering distance cutoff value, passed on as parameter h to cutree. Note that if this is smaller than the smallest nonzero geographical distance, communities will be all sets of individuals that have zero geographical distance to each other.
method	method-parameter for hclust.

Value

Vector of community memberships for the individuals (integer numbers from 1 to the number of communities without interruption.

Author(s)

Christian Hennig <christian.hennig@unibo.it> https://www.unibo.it/sitoweb/christian.hennig/en

See Also

communitydist

Examples

```
data(veronica)
ver.geo <- coord2dist(coordmatrix=veronica.coord[1:90,],file.format="decimal2")
species <-c(rep(1,64),rep(2,17),rep(3,9))
communities(ver.geo,species)
```

communitydist *Distances between communities*

Description

Constructs distances between communities: chord- (Cavalli-Sforza and Edwards, 1967), phiPT/phiST (Peakall and Smouse, 2012, Meirmans, 2006), three versions of the shared allele distance between communities, and geographical distance between communities.

Usage

```
communitydist(alleleobj,comvector="auto",distance="chord",
                          compute.geodist=TRUE,out.dist=FALSE,
                          grouping=NULL,geodist=NA,diploid=TRUE,
                          phiptna=NA,...)
```

Arguments

alleleobj	if diploid=TRUE, an object of class alleleobject as produced by function alleleinit. This has the required information on the individuals that are grouped into communities. In case diploid=FALSE, a list that needs to have components n.variables (number of loci), alevels (vector of allele names, see alleleinit) and charmatrix (matrix of characters with one row for every individual and one column for every locus giving the alleles; see examples below for how this can be constructed for a prabobject with presence-absence data).
comvector	either a vector of integers indicating to which community an individual belongs (these need to be numbered from 1 to a maximum number without interruption), or "auto", which indicates that communities are automatically generated by the communities-function.
distance	one of "chord", "phipt", "shared.average", "shared.chakraborty", "shared.problist". See Details.
compute.geodist	logical, indicating whether geographical distances between communities should be generated.
out.dist	logical, indicating whether dist-objects are given out or rather distance matrices.
grouping	something that can be coerced into a factor, for passing on to communities in case that comvector=="auto". This implies that individuals in different groups indicated by grouping cannot be together in the same community. Furthermore (also if comvector is something else), a vector of groups of communities will be computed, see output component comgroup. In any case individuals in different groups are not allowed to be in the same community.

geodist	matrix or dist-object providing geographical distances between individuals. Required if compute.geodist==TRUE or comvector=="auto".
diploid	logical, indicating whether loci are diploid, see alleleobj.
phiptna	if distance="phipt", value to be given out as phiPT-distance in case that the original definition amounts to 0/0 (particularly if communities have just one member).
...	optional arguments to be passed on to communities.

Details

All genetic distances between communities are based on the information given in alleleobj; either on the alleles directly or on a genetic distance (distmat-component, see alleleinit). The possible genetic distance measures between communities are as follows:

- "chord": chord-distance (Cavalli-Sforza and Edwards, 1967)
- "phipt": phiPT-distance implemented according to Peakall and Smouse, 2012. This also appears in the literature under the name phiST (Meirmans, 2006, although the definition there is incomplete and we are not sure whether this is identical).
- "shared.average": average of between-community genetic distances.
- "shared.chakraborty": between-community shared allele distance according to Chakraborty and Jin (1993).
- "shared.problist": this implements the shared allele distance (Bowcock et al., 1994) for individuals directly for communities (one minus proportion of alleles shared by two communities averaged over loci).

Value

list with components

comvector	integer vector of length of the number of individuals, indicating their community membership.
dist	genetic distances between communities. Parameter out.dist determines whether this is a dist-object or a matrix.
cgeodist	if compute.geodist, geographical distance between communities defined as average distance of all pairs of individuals belonging to different ones of the two communities between which the distance is computed. Parameter out.dist determines whether this is a dist-object or a matrix.
comgroup	vector of length of the number of communities. If grouping was provided, this is a vector giving the group memberships of all communities, otherwise it is a vector of 1s.

Author(s)

Christian Hennig <christian.hennig@unibo.it> https://www.unibo.it/sitoweb/christian.hennig/en

References

Bowcock, A. M., Ruiz-Linares, A., Tomfohrde, J., Minch, E., Kidd, J. R., Cavalli-Sforza, L. L. (1994) High resolution of human evolutionary trees with polymorphic microsatellites. *Nature* 368, 455-457.

Cavalli-Sforza, L. L. and Edwards, A. W. F. (1967) Phylogenetic Analysis - Models and Estimation Procedures. *The American Journal of Human Genetics* 19, 233-257.

Chakraborty, R. and Jin, L. (1993) Determination of relatedness between individuals using DNA fingerprinting. *Human Biology* 65, 875-895.

Meirmans, P. G. (2006) Using the AMOVA framework to estimate a standardized genetic differentiation measure. *Evolution* 60, 2399-2402.

Peakall, R. and Smouse P.E. (2012) GenAlEx Tutorial 2. https://biology-assets.anu.edu.au/GenAlEx/Tutorials.html

See Also

communities; refer to phipt for computation of distances between specific pairs of communities. diploidcomlist produces relative frequencies for all alles of all loci in all communities (on which the chord- and the "shared.problist"-distances are based).

Examples

```
options(digits=4)
data(tetragonula)
tnb <-
coord2dist(coordmatrix=tetragonula.coord[83:120,],cut=50,
  file.format="decimal2",neighbors=TRUE)
ta <- alleleconvert(strmatrix=tetragonula[83:120,])
tai <- alleleinit(allelematrix=ta,neighborhood=tnb$nblist)
tetraspec <- c(rep(1,11),rep(2,13),rep(3,14))
tetracoms <-
c(rep(1:3,each=3),4,5,rep(6:11,each=2),12,rep(13:19,each=2))
c1 <- communitydist(tai,comvector=tetracoms,distance="chord",
  geodist=tnb$distmatrix,grouping=tetraspec)
c2 <- communitydist(tai,comvector=tetracoms,distance="phipt",
  geodist=tnb$distmatrix,grouping=tetraspec,compute.geodist=FALSE)
c3 <- communitydist(tai,comvector=tetracoms,distance="shared.average",
  geodist=tnb$distmatrix,grouping=tetraspec,compute.geodist=FALSE)
c4 <- communitydist(tai,comvector=tetracoms,distance="shared.chakraborty",
  geodist=tnb$distmatrix,grouping=tetraspec,compute.geodist=FALSE)
c5 <- communitydist(tai,comvector=tetracoms,distance="shared.problist",
  geodist=tnb$distmatrix,grouping=tetraspec,compute.geodist=FALSE)
round(c1$cgeodist,digits=1)
c1$comvector
c2$comvector
c3$comvector
c4$comvector
c5$comvector
round(c1$dist,digits=2)
round(c2$dist,digits=2)
```

```
round(c3$dist,digits=2)
round(c4$dist,digits=2)
round(c5$dist,digits=2)
```

`comp.test` *Compare species clustering and species groups*

Description

Tests for independence between a clustering and another grouping of species. This is simply an interface to `chisq.test`.

Usage

```
comp.test(cl,spg)
```

Arguments

`cl`	a vector of integers. Clustering of species (may be taken from `prabclust`).
`spg`	a vector of integers of the same length, groups of species.

Details

`chisq.test` with simulated p-value is used.

Value

Output of `chisq.test`.

Author(s)

Christian Hennig <christian.hennig@unibo.it> https://www.unibo.it/sitoweb/christian.hennig/en

References

Hausdorf, B. and Hennig, C. (2003) Biotic Element Analysis in Biogeography. *Systematic Biology* 52, 717-723.

See Also

`chisq.test`, `prabclust`.

Examples

```
set.seed(1234)
g1 <- c(rep(1,34),rep(2,12),rep(3,15))
g2 <- sample(3,61,replace=TRUE)
comp.test(g1,g2)
```

con.comp *Connectivity components of an undirected graph*

Description

Computes the connectivity components of an undirected graph from a matrix giving the edges.

Usage

```
con.comp(comat)
```

Arguments

comat	a symmetric logical or 0-1 matrix, where comat[i,j]=TRUE means that there is an edge between vertices i and j. The diagonal is ignored.

Details

The "depth-first search" algorithm of Cormen, Leiserson and Rivest (1990, p. 477) is used.

Value

An integer vector, giving the number of the connectivity component for each vertice.

Author(s)

Christian Hennig <christian.hennig@unibo.it> https://www.unibo.it/sitoweb/christian.hennig/en

References

Cormen, T. H., Leiserson, C. E. and Rivest, R. L. (1990), *Introduction to Algorithms*, Cambridge: MIT Press.

See Also

hclust, cutree for cutted single linkage trees (often equivalent).

Examples

```
set.seed(1000)
x <- rnorm(20)
m <- matrix(0,nrow=20,ncol=20)
for(i in 1:20)
  for(j in 1:20)
    m[i,j] <- abs(x[i]-x[j])
d <- m<0.2
cc <- con.comp(d)
max(cc) # number of connectivity components
```

```
plot(x,cc)
# The same should be produced by
# cutree(hclust(as.dist(m),method="single"),h=0.2).
```

---

con.regmat *Connected regions per species*

Description

Returns a vector of the numbers of connected regions per species for a presence-absence matrix.

Usage

```
con.regmat(regmat, neighbors, count = FALSE)
```

Arguments

regmat	0-1-matrix. Columns are species, rows are regions.
neighbors	A list with a component for every region. The components are vectors of integers indicating neighboring regions. A region without neighbors (e.g., an island) should be assigned a list `numeric(0)`.
count	logical. If `TRUE`, the number of the currently processed species is printed.

Details

Uses `con.comp`.

Value

Vector of numbers of connected regions per species.

Note

Designed for use in `prabtest`.

Author(s)

Christian Hennig <christian.hennig@unibo.it> https://www.unibo.it/sitoweb/christian.hennig/en

See Also

con.comp, prabtest

Examples

```
data(nb)
set.seed(888)
cp <- cluspop.nb(nb, p.nb=0.1, n.species=10, clus.specs=9,
                 reg.group=1:17,vector.species=c(10))
con.regmat(cp,nb)
```

`coord2dist` *Geographical coordinates to distances*

Description

Computes geographical distances from geographical coordinates

Usage

```
coord2dist(file=NULL, coordmatrix=NULL, cut=NULL,
                    file.format="degminsec",
                    output.dist=FALSE, radius=6378.137,
            fp=1/298.257223563, neighbors=FALSE)
```

Arguments

`file`	string. A filename for the coordinate file. The file should have 2, 4 or 6 numeric columns and one row for each location. See `file.format`. One of `file` and `coordmatrix` needs to be specified (if `coordmatrix` is not specified, coordinates are read from `file`).
`coordmatrix`	something that can be coerced into a matrix with 2, 4 or 6 columns. Matrix of coordinates, one row for each location. See `file.format`. One of `file` and `coordmatrix` needs to be specified.
`cut`	numeric. Only active if `neighbors==TRUE`; see `neighbors`.
`file.format`	one of `"degminsec"`, `"decimal2"` or `"decimal4"`. The format of the required file or `coordmatrix` consists of the following columns:

"degminsec" 6 columns; the first three give degrees, minutes and seconds for latitude, columns 4-6 the same for longitude. Values in column 1 and 4 can be positive or negative (negative means "South", "West", respectively). Values in the other columns should be non-negative.

"decimal2" 2 columns; the first one gives latitude, the second one longitude in proper decimal notation. Values can be positive or negative (negative means "South", "West", respectively).

"decimal4" 4 columns; the first two give latitude, no. 3 and 4 give longitude. Values in column 1 and 3 can be positive or negative (negative means "South", "West", respectively). The give integer degrees. Values in the other columns should be non-negative. They give percentages (`<=100`).

output.dist	logical. If TRUE, the resulting distance matrix is given out as a dist object.
radius	numeric. Radius of the earth in km used in computation (the default is the equatorial radius but this is not the uniquely possible choice).
fp	flattening of the earth; the default is from WGS-84.
neighbors	logical. If TRUE, a neighborhood list is also computed, listing for every location all locations with distance <=cut as neighbors.

Value

If neighbors==TRUE, a list with components

distmatrix	distance matrix between locations. See output.dist above. This is in km by default; the measurement unit is determined by the value used for radius.
nblist	list with a vector for every location containing the numbers of its neighbors, see neighbors.

If neighbors==FALSE, only the distance matrix.

Author(s)

Christian Hennig <christian.hennig@unibo.it> https://www.unibo.it/sitoweb/christian.hennig/en

References

German Wikipedia from 29 August 2010: https://de.wikipedia.org/wiki/Orthodrome

See Also

geo2neighbor

Examples

```
options(digits=4)
data(veronica)
coord2dist(coordmatrix=veronica.coord[1:20,], cut=20, file.format="decimal2",neighbors=TRUE)
```

---

crmatrix *Region-wise cluster membership*

Description

Produces a matrix with clusters as rows and regions as columns, indicating how many species present in a region belong to the clusters

Usage

```
crmatrix(x,xc,percentages=FALSE)
```

Arguments

x	object of class prab as generated by prabinit. Presence-absence data to be analyzed.
xc	object of class prabclust or comprabclust as generated by prabclust or hprabclust. The clustering.
percentages	logical. If TRUE, the output matrix will give the proportion of species from a certain region in the cluster.

Value

A clusters time regions matrix as explained above.

Author(s)

Christian Hennig <christian.hennig@unibo.it> https://www.unibo.it/sitoweb/christian.hennig/en

Examples

```
options(digits=3)
data(kykladspecreg)
data(nb)
set.seed(1234)
x <- prabinit(prabmatrix=kykladspecreg, neighborhood=nb)
xc <- prabclust(x)

crmatrix(x,xc)
crmatrix(x,xc, percentages=TRUE)
```

---

dicedist *Dice distance matrix*

Description

Computes a distance derived from Dice's coincidence index between the columns of a 0-1-matrix.

Usage

```
dicedist(regmat)
```

Arguments

regmat	0-1-matrix. Columns are species, rows are regions.

Details

The Dice distance between two species is 1 minus the Coincidence Index, which is (2*number of regions where both species are present)/(2*number of regions where both species are present plus number of regions where at least one species is present). This is S23 in Shi (1993).

Value

A symmetrical matrix of Dice distances.

Author(s)

Christian Hennig <christian.hennig@unibo.it> https://www.unibo.it/sitoweb/christian.hennig/en

References

Shi, G. R. (1993) Multivariate data analysis in palaeoecology and palaeobiogeography - a review. *Palaeogeography, Palaeoclimatology, Palaeoecology* 105, 199-234.

See Also

kulczynski,jaccard

Examples

```
options(digits=4)
data(kykladspecreg)
dicedist(t(kykladspecreg))
```

---

distratio *Distance ratio test statistics for distance based clustering*

Description

Calculates the ratio between the prop smallest and largest distances of a distance matrix.

Usage

```
distratio(distmat, prop = 0.25)
```

Arguments

distmat	symmetric distance matrix.
prop	numerical. Proportion between 0 and 1.

Details

Rounding is by floor for small and ceiling for large distances.

Value

A list with components

dr	ratio of prop smallest to prop largest distances.
lowmean	mean of prop smallest distances.
himean	mean of prop smallest distances.
prop	see above.

Author(s)

Christian Hennig <christian.hennig@unibo.it> https://www.unibo.it/sitoweb/christian.hennig/en

References

Hennig, C. and Hausdorf, B. (2004) Distance-based parametric bootstrap tests for clustering of species ranges. *Computational Statistics and Data Analysis* 45, 875-896.

See Also

prabtest

Examples

```
options(digits=4)
data(kykladspecreg)
j <- jaccard(t(kykladspecreg))
distratio(j)
```

---

geco *geco distance matrix*

Description

Computes geco distances between the columns of a 0-1-matrix, based on a distance matrix between regions (usually, but not necessarily, this is a geographical distance).

Usage

```
geco(regmat,geodist=as.dist(matrix(as.integer(!diag(nrow(regmat)))),
                    transform="piece",
                    tf=0.1,
                    countmode=ncol(regmat)+1)
```

Arguments

regmat	0-1-matrix. Columns are species, rows are regions.
geodist	dist-object or symmetric non-negative matrix. Geographical distances between regions.
transform	transformation applied to the distances before computation of geco coefficient, see details. "piece" means piecewise linear, namely distance/(tf*maximum distance) if distance<tf*maximum distance, and 1 otherwise, "log" means log((tf*distance)+1), "sqrt" means sqrt(tf*distance), "none" means no transformation.
tf	tuning constant for transformation. See transform.
countmode	optional positive integer. Every 'countmode' algorithm runs 'geco' shows a message.

Details

The geco distance between two species is 0.5*(mean distance between region where species 1 is present and closest region where species 2 is present plus mean distance between region where species 2 is present and closest region where species 1 is present). 'closest' to a region could be the regions itself. It is recommended (Hennig and Hausdorf, 2006) to transform the distances first, because the differences between large distances are usually not meaningful or at least much less meaningful than differences between small distances for dissimilarity measurement between species ranges. See parameter transform.

If the between-regions distance is 1 for all pairs of non-equal regions, the geco distance degenerates to the Kulczynski distance, see kulczynski.

Value

A symmetrical matrix of geco distances.

Author(s)

Christian Hennig <christian.hennig@unibo.it> https://www.unibo.it/sitoweb/christian.hennig/en

References

Hennig, C. and Hausdorf, B. (2006) A robust distance coefficient between distribution areas incorporating geographic distances. *Systematic Biology* 55, 170-175.

See Also

kulczynski

Examples

```
options(digits=4)
data(kykladspecreg)
data(waterdist)
geco(t(kykladspecreg),waterdist)
```

geo2neighbor *Neighborhood list from geographical distance*

Description

Generates a neighborhood list as required by `prabinit` from a matrix of geographical distances.

Usage

```
geo2neighbor(geodist,cut=0.1*max(geodist))
```

Arguments

`geodist`	`dist`-object or symmetric non-negative matrix. Geographical distances between regions.
`cut`	non-negative numerical. All pairs of regions with `distance<=cut` are treated as neighbors.

Value

A list of integer vectors, giving the set of neighbors for every region.

Author(s)

Christian Hennig <christian.hennig@unibo.it> https://www.unibo.it/sitoweb/christian.hennig/en

Examples

```
data(waterdist)
geo2neighbor(waterdist)
```

homogen.test *Classical distance-based test for homogeneity against clustering*

Description

Classical distance-based test for homogeneity against clustering. Test statistics is number of isolated vertices in the graph of smallest distances. The homogeneity model is a random graph model where `ne` edges are drawn from all possible edges.

Usage

```
homogen.test(distmat, ne = ncol(distmat), testdist = "erdos")
```

Arguments

distmat	numeric symmetric distance matrix.
ne	integer. Number of edges in the data graph, corresponding to smallest distances.
testdist	string. If testdist="erdos", the test distribution is a Poisson asymptotic distibution as given by Erdos and Renyi (1960). If testdist="ling", the test distribution is exact as given by Ling (1973), which needs much more computing time.

Details

The "ling"-test is one-sided (rejection if the number of isolated vertices is too large), the "erdos"-test computes a one-sided as well as a two-sided p-value.

Value

A list with components

p	p-value for one-sided test.
p.twoside	p-value for two-sided test, only if testdist="erdos".
iv	number of isolated vertices in the data.
lambda	parameter of the Poisson test distribution, only if testdist="erdos".
distcut	largest distance value for which an edge has been drawn.
ne	see above.

Author(s)

Christian Hennig <christian.hennig@unibo.it> https://www.unibo.it/sitoweb/christian.hennig/en

References

Erdos, P. and Renyi, A. (1960) On the evolution of random graphs. *Publications of the Mathematical Institute of the Hungarian Academy of Sciences* 5, 17-61.

Godehardt, E. and Horsch, A. (1995) Graph-Theoretic Models for Testing the Homogeneity of Data. In Gaul, W. and Pfeifer, D. (Eds.) *From Data to Knowledge*, Springer, Berlin, 167-176.

Ling, R. F. (1973) A probability theory of cluster analysis. *Journal of the American Statistical Association* 68, 159-164.

See Also

prabtest

Examples

```
options(digits=4)
data(kykladspecreg)
j <- jaccard(t(kykladspecreg))
homogen.test(j, testdist="erdos")
homogen.test(j, testdist="ling")
```

`hprabclust` *Clustering of species ranges from presence-absence matrices (hierarchical methods)*

Description

Clusters a presence-absence matrix object by taking the 'h-cut'-partition of a hierarchical clustering and declaring all members of too small clusters as 'noise' (this gives a distance-based clustering method, which estimates the number of clusters and allows for noise/non-clustered points). Note that this is experimental. Often, the `prabclust`-solutions is more convincing due to higher flexibility of that method. However, `hprabclust` may be more stable sometimes.

Note: Data formats are described on the `prabinit` help page. You may also consider the example datasets `kykladspecreg.dat` and `nb.dat`. Take care of the parameter `rows.are.species` of `prabinit`.

Usage

```
hprabclust(prabobj, cutdist=0.4, cutout=1,
method="average", nnout=2, mdsplot=TRUE, mdsmethod="classical")

## S3 method for class 'comprabclust'
print(x, ...)
```

Arguments

`prabobj`	object of class `prab` as generated by `prabinit`. Presence-absence data to be analyzed.
`cutdist`	non-negative integer. Cutoff distance to determine the partition, see `cutree`.
`cutout`	non-negative integer. Points that have at most `nnout` distances smaller or equal than `cutout` are treated as noise.
`method`	string. Clustering method, see `hclust`.
`nnout`	non-negative integer. Members of clusters with less or equal than `nnout` points or that have less or equal than `nnout` neighbors closer than `cutout` are treated as noise.
`mdsplot`	logical. If `TRUE`, the cluster solution is plotted on the first two MDS dimensions, see `mdsmethod`.

mdsmethod	"classical", "kruskal", or "sammon". The MDS method to transform the distances to data points. "classical" indicates metric MDS by function cmdscale, "kruskal" is non-metric MDS. Note that if mdsmethod!="classical" zero distances between different objects are replaced by the minimum of the nonzero distances divided by 10 (otherwise the MDS method would produce an error). Note that mdsmethod is ignored if mdsplot=FALSE.
x	comprabclust-object as generated by hprabclus.
...	necessary for print method.

Value

hprabclust generates an object of class comprabclust. This is a list with components

clustering	vector of integers indicating the cluster memberships of the species (cutout-outliers are noise, but small clusters are allowed). Noise is coded as 0.
rclustering	vector of integers indicating the cluster memberships of the species, noise as described under nnout. Noise is coded as 0.
cutdist	see above.
method	see above.
cutout	see above.
nnout	see above.
noisen	number of points minus cutout-outliers.
symbols	vector of characters corresponding to rclustering, but estimated noise by "N".
points	numerical matrix. MDS configuration (if mdsplot=TRUE).
call	function call.

Author(s)

Christian Hennig <christian.hennig@unibo.it> https://www.unibo.it/sitoweb/christian.hennig/en

See Also

hclust, cutree, prabclust.

Examples

```
data(kykladspecreg)
data(nb)
data(waterdist)
x <- prabinit(prabmatrix=kykladspecreg, neighborhood=nb,
              geodist=waterdist, distance="geco")
hprabclust(x,mdsplot=FALSE)
```

incmatrix *Nestedness matrix*

Description

Computes species*species nestedness matrix and number of nestings (inclusions) from regions*species presence-absence matrix.

Usage

```
incmatrix(regmat)
```

Arguments

regmat	0-1-matrix. Columns are species, rows are regions.

Value

A list with components

m	0-1-matrix. m[i,j]=1 means that the occupied region of species j is a subset (not equal) of the region of species i.
ninc	integer. Number of strict inclusions.
neq	integer. Number of region equalities between species (not including equality between species i and i).

Author(s)

Christian Hennig <christian.hennig@unibo.it> https://www.unibo.it/sitoweb/christian.hennig/en

References

Hausdorf, B. and Hennig, C. (2003) Nestedness of nerth-west European land snail ranges as a consequence of differential immigration from Pleistocene glacial refuges. *Oecologia* 135, 102-109.

See Also

prabtest

Examples

```
data(kykladspecreg)
incmatrix(t(kykladspecreg))$ninc
```

jaccard *Jaccard distance matrix*

Description

Computes Jaccard distances between the columns of a 0-1-matrix.

Usage

```
jaccard(regmat)
```

Arguments

regmat	0-1-matrix. Columns are species, rows are regions.

Details

The Jaccard distance between two species is 1-(number of regions where both species are present)/(number of regions where at least one species is present). As a similarity coefficient, this is S22 in Shi (1993).

Thank you to Laurent Buffat for improving this function!

Value

A symmetrical matrix of Jaccard distances.

Author(s)

Christian Hennig <christian.hennig@unibo.it> https://www.unibo.it/sitoweb/christian.hennig/en

References

Shi, G. R. (1993) Multivariate data analysis in palaeoecology and palaeobiogeography - a review. *Palaeogeography, Palaeoclimatology, Palaeoecology* 105, 199-234.

See Also

kulczynski, dicedist

Examples

```
options(digits=4)
data(kykladspecreg)
jaccard(t(kykladspecreg))
```

kulczynski *Kulczynski distance matrix*

Description

Computes Kulczynski distances between the columns of a 0-1-matrix.

Usage

```
kulczynski(regmat)
```

Arguments

regmat	0-1-matrix. Columns are species, rows are regions.

Details

The Kulczynski distance between two species is 1-(mean of (number of regions where both species are present)/(number of regions where species 1 is present) and (number of regions where both species are present)/(number of regions where species 2 is present)). The similarity version of this is S28 in Shi (1993).

Value

A symmetrical matrix of Kulczynski distances.

Author(s)

Christian Hennig <christian.hennig@unibo.it> https://www.unibo.it/sitoweb/christian.hennig/en

References

Shi, G. R. (1993) Multivariate data analysis in palaeoecology and palaeobiogeography - a review. *Palaeogeography, Palaeoclimatology, Palaeoecology* 105, 199-234.

See Also

jaccard, geco,qkulczynski , dicedist

Examples

```
options(digits=4)
data(kykladspecreg)
kulczynski(t(kykladspecreg))
```

kykladspecreg *Snail presence-absence data from Aegean sea*

Description

0-1-matrix where rows are snail species and columns are islands in the Aegean sea. An entry of 1 means that the species is present in the region.

Usage

```
data(kykladspecreg)
```

Format

A 0-1 matrix with 80 rows and 34 columns.

Details

Reads from example data file `kykladspecreg.dat`.

Source

B. Hausdorf and C. Hennig (2005) The influence of recent geography, palaeography and climate on the composition of the faune of the central Aegean Islands. *Biological Journal of the Linnean Society* 84, 785-795.

See Also

`nb` provides neighborhood information about the 34 islands. `waterdist` provides a geographical distance matrix between the islands.

Examples

```
data(kykladspecreg)
```

lcomponent *Largest connectivity component*

Description

Computes the size of the largest connectivity component of the graph of `ncol(distmat)` vertices with edges defined by the smallest `ne` distances.

Usage

```
lcomponent(distmat, ne = floor(3*ncol(distmat)/4))
```

Arguments

distmat	symmetric distance matrix.
ne	integer.

Value

list with components

lc	size of the largest connectivity component.
ne	see above.

Author(s)

Christian Hennig <christian.hennig@unibo.it> https://www.unibo.it/sitoweb/christian.hennig/en

References

Hennig, C. and Hausdorf, B. (2004) Distance-based parametric bootstrap tests for clustering of species ranges. *Computational Statistics and Data Analysis* 45, 875-896.

See Also

prabtest

Examples

```
data(kykladspecreg)
j <- jaccard(t(kykladspecreg))
lcomponent(j)
```

---

lociplots *Visualises clusters of markers vs. species*

Description

Given a clustering of individuals from prabclust (as generated in species delimitation) and a clustering of markers (for example dominant markers of genetic loci), lociplots visualises the presence of markers against the clustering of individuals and computes some statistics.

Usage

```
lociplots(indclust,locclust,locprab,lcluster,
                    symbols=NULL,brightest.grey=0.8,darkest.grey=0,
                    mdsdim=1:2)
```

Arguments

indclust	prabclust-object. Clustering of individuals.
locclust	vector of integers. Clustering of markers/loci.
locprab	prab-object in which the markers are what the help page of prabinit refers to as "species" (i.e., reverse of what is used for species delimitation clustering; for data sets with codominant markers, such an object can be constructed by use of allele2zeroone before prabinit.)
lcluster	integer. Number of cluster in locclust for which plot and statistics are produced.
symbols	vector of plot symbols. If NULL, indclust$symbols is used.
brightest.grey	numeric between 0 and 1. Brightest grey value used in plot for individuals with smallest marker percentage, see details.
darkest.grey	numeric between 0 and 1. Darkest grey value used in plot for individuals with highest marker percentage, see details.
mdsdim	vector of two integers. The two MDS variables taken from indclust used for visualisation.

Details

Plot and statistics are based on the individual marker percentage, which is the percentage of markers present in an individual of the markers belonging to cluster no. lcluster. In the plot, the grey value visualises the marker percentage.

Value

list with components

locfreq	vector of individual marker percentages.
locfreqmin	vector of minimum individual marker precentages for each cluster in indclust-clustering (the first value refers to the "noise component", if present).
locfreqmax	vector of maximum individual marker precentages for each cluster in indclust-clustering (the first value refers to the "noise component", if present).
locfreqmean	vector of average individual marker precentages for each cluster in indclust-clustering (the first value refers to the "noise component", if present).

Author(s)

Christian Hennig <christian.hennig@unibo.it> https://www.unibo.it/sitoweb/christian.hennig/en

See Also

prabclust

Examples

```
options(digits=4)
data(veronica)
vei <- prabinit(prabmatrix=veronica[1:50,],distance="jaccard")
ppv <- prabclust(vei)
veloci <- prabinit(prabmatrix=veronica[1:50,],rows.are.species=FALSE)
velociclust <- prabclust(veloci,nnk=0)
lociplots(ppv,velociclust$clustering,veloci,lcluster=3)
```

`nastats` *Missing values statistics for matrix*

Description

Computes column-wise and row-wise numbers of missing values.

Usage

```
nastats(amatrix, nastr="--")
```

Arguments

`amatrix`	(any) matrix.
`nastr`	missing value indicator.

Value

A list with components

`narow`	vector of row-wise numbers of mixxing values.
`nacol`	vector of column-wise numbers of mixxing values.

Author(s)

Christian Hennig <christian.hennig@unibo.it> https://www.unibo.it/sitoweb/christian.hennig/en

Examples

```
xx <- cbind(c(1,2,3),c(0,0,1),c(5,3,1))
nastats(xx,nastr=0)
```

nb *Neighborhood list for Aegean islands*

Description

List of neighboring islands for 34 Aegean islands.

Usage

```
data(nb)
```

Format

List with 34 components, all being vetors of integers (or `numeric(0)` in case of no neighbors) indicating the neighboring islands.

Details

Reads from example data file `nb.dat`.

Source

B. Hausdorf and C. Hennig (2005) The influence of recent geography, palaeography and climate on the composition of the faune of the central Aegean Islands. *Biological Journal of the Linnean Society* 84, 785-795.

Examples

```
data(nb)
# nb <- list()
# for (i in 1:34)
#   nb <- c(nb,list(scan(file="(path/)nb.dat",
#                   skip=i-1,nlines=1)))
```

nbtest *Test of neighborhood list*

Description

Tests a list of neighboring regions for proper format. Neighborhood is tested for being symmetrical. Causes an error if tests fail.

Usage

```
nbtest(nblist, n.regions=length(nblist))
```

Arguments

nblist	A list with a component for every region. The components are vectors of integers indicating neighboring regions. A region without neighbors (e.g., an island) should be assigned a vector numeric(0).
n.regions	Number of regions.

Value

invisible{TRUE}.

Author(s)

Christian Hennig <christian.hennig@unibo.it> https://www.unibo.it/sitoweb/christian.hennig/en

See Also

prabinit.

Examples

```
data(nb)
nbtest(nb)
nb[[1]][1] <- 1
try(nbtest(nb))
```

---

nn *Mean distance to kth nearest neighbor*

Description

Computes the mean of the distances from each point to its neth nearest neighbor.

Usage

```
nn(distmat, ne = 1)
```

Arguments

distmat	symmetric distance matrix (not a dist-object).
ne	integer.

Value

numerical.

Author(s)

Christian Hennig <christian.hennig@unibo.it> https://www.unibo.it/sitoweb/christian.hennig/en

References

Hennig, C. and Hausdorf, B. (2004) Distance-based parametric bootstrap tests for clustering of species ranges. *Computational Statistics and Data Analysis* 45, 875-896.

See Also

prabtest

Examples

```
data(kykladspecreg)
j <- jaccard(t(kykladspecreg))
nn(j,4)
```

NNclean *Nearest neighbor based clutter/noise detection*

Description

Detects if data points are noise or part of a cluster, based on a Poisson process model.

Usage

```
NNclean(data, k, distances = NULL, edge.correct = FALSE, wrap = 0.1,
convergence = 0.001, plot=FALSE, quiet=TRUE)

## S3 method for class 'nnclean'
print(x, ...)
```

Arguments

data	numerical matrix or data frame.
k	integer. Number of considered nearest neighbors per point.
distances	distance matrix object of class dist. If specified, it is used instead of computing distances from the data.
edge.correct	logical. If TRUE and the data is two-dimensional, neighbors for points at the edges of the parent region of the noise Poisson process are determined after wrapping the region onto a toroid.
wrap	numerical. If edge.correct=TRUE, points in a strip of size wrap*range along the edge for each variable are candidates for being neighbors of points from the opposite.

convergence	numerical. Convergence criterion for EM-algorithm.
plot	logical. If TRUE, a histogram of the distance to kth nearest neighbor and fit is plotted.
quiet	logical. If FALSE, the likelihood is printed during the iterations.
x	object of class nnclean.
...	necessary for print methods.

Details

The assumption is that the noise is distributed as a homogeneous Poisson process on a certain region and the clusters are distributed as a homogeneous Poisson process with larger intensity on a subregion (disconnected in case of more than one cluster). The distances are then distributed according to a mixture of two transformed Gamma distributions, and this mixture is estimated via the EM-algorithm. The points are assigned to noise or cluster component by use of the estimated a posteriori probabilities.

Value

NNclean returns a list of class nnclean with components

z	0-1-vector of length of the number of data points. 1 means cluster, 0 means noise.
probs	vector of estimated a priori probabilities for each point to belong to the cluster component.
k	see above.
lambda1	intensity parameter of cluster component.
lambda2	intensity parameter of noise component.
p	estimated probability of cluster component.
kthNND	distance to kth nearest neighbor.

Note

The software can be freely used for non-commercial purposes, and can be freely distributed for non-commercial purposes only.

Author(s)

R-port by Christian Hennig <christian.hennig@unibo.it> https://www.unibo.it/sitoweb/christian.hennig/en,
original Splus package by S. Byers and A. E. Raftery.

References

Byers, S. and Raftery, A. E. (1998) Nearest-Neighbor Clutter Removal for Estimating Features in Spatial Point Processes, *Journal of the American Statistical Association*, 93, 577-584.

Examples

```
library(mclust)
data(chevron)
nnc <-  NNclean(chevron[,2:3],15,plot=TRUE)
plot(chevron[,2:3],col=1+nnc$z)
```

---

phipt *Distances between communities, auxiliary functions*

Description

Auxiliary functions for communitydist. phipt computes phiPT/phiST (Peakall and Smouse, 2012, Meirmans, 2006) between two communities. cfchord computes the chord-distance (Cavalli-Sforza and Edwards, 1967) between two lists or locus-wise relative allele frequencies. shared.problist computes a straightforward generalisation of the shared allele distance (Bowcock et al., 1994) between individuals for communities, namely the 'overlap', i.e., sum of the minima of the allele relative frequencies. diploidcomlist constructs the input lists for cfchord and shared.problist from an alleleobject. It provides relative frequencies for all alles of all loci in all communities.

Usage

```
phipt(alleleobj,comvector,i,j)
cfchord(p1,p2)
shared.problist(p1,p2)
diploidcomlist(alleleobj,comvector,diploid=TRUE)
```

Arguments

alleleobj	if diploid=TRUE, an object of class alleleobject as produced by function alleleinit. This has the required information on the individuals that are grouped into communities. In case diploid=FALSE, a list that needs to have components n.variables (number of loci), alevels (vector of allele names, see alleleinit) and charmatrix (matrix of characters with one row for every individual and one column for every locus giving the alleles; see examples below for how this can be constructed for a prabobject with presence-absence data).
comvector	vector of integers indicating to which community an individual belongs.
i	integer. Number of community.
j	integer. Number of community. The phiPT-distance is computed between the communities numbered i and j
p1	list. Every list entry refers to a locus and is a vector of relative frequencies of the alleles present in that locus in a community.
p2	list. Every list entry refers to a locus and is a vector of relative frequencies of the alleles present in that locus in a community. The chord or shared allele distance is computed between the communities encoded by p1 and p2.
diploid	logical, indicating whether loci are diploid, see alleleobj.

Value

`cfchord` gives out the value of the chord distance. `shared.problist` gives out the distance value. `diploidcomlist` gives out a two-dimensional list. The list has one entry for each community, which is itself a list. This community list has one entry for each locus, which is a vector that gives the relative frequencies of the different alleles in `phipt` gives out a list with components `phipt`, `vap`, `n0`, `sst`, `ssg`, `msa`, `msw`. These refer to the notation on p.2.12 and 2.15 of Peakall and Smouse (2012).

`phipt`	value of phiPT.
`vap`	variance among (between) populations (communities).
`n0`	standardisation factor N0, see p.2.12 of Peakall and Smouse (2012).
`sst`	total distances sum of squares.
`ssg`	vector with two non-`NA` entriesm, within community sums of squares for communities `i` and `j`.
`msa`	mean square between communities.
`msw`	mean square within communities.

Author(s)

Christian Hennig <christian.hennig@unibo.it> https://www.unibo.it/sitoweb/christian.hennig/en

References

Bowcock, A. M., Ruiz-Linares, A., Tomfohrde, J., Minch, E., Kidd, J. R., Cavalli-Sforza, L. L. (1994) High resolution of human evolutionary trees with polymorphic microsatellites. *Nature* 368, 455-457.

Cavalli-Sforza, L. L. and Edwards, A. W. F. (1967) Phylogenetic Analysis - Models and Estimation Procedures. *The American Journal of Human Genetics* 19, 233-257.

Meirmans, P. G. (2006) Using the AMOVA framework to estimate a standardized genetic differentiation measure. *Evolution* 60, 2399-2402.

Peakall, R. and Smouse P.E. (2012) GenAlEx Tutorial 2. https://biology-assets.anu.edu.au/GenAlEx/Tutorials.html

See Also

`communitydist`

Examples

```
options(digits=4)
data(tetragonula)
tnb <-
coord2dist(coordmatrix=tetragonula.coord[83:120,],cut=50,file.format="decimal2",neighbors=TRUE)
ta <- alleleconvert(strmatrix=tetragonula[83:120,])
tai <- alleleinit(allelematrix=ta,neighborhood=tnb$nblist)
tetracoms <-
```

```
c(rep(1:3,each=3),4,5,rep(6:11,each=2),12,rep(13:19,each=2))
phipt(tai,tetracoms,4,6)
tdip <- diploidcomlist(tai,tetracoms,diploid=TRUE)
cfchord(tdip[[4]],tdip[[6]])
shared.problist(tdip[[4]],tdip[[6]])
```

`piecewiselin` *Piecewise linear transformation for distance matrices*

Description

Piecewise linear transformation for distance matrices, utility function for geco.

Usage

```
piecewiselin(distmatrix, maxdist=0.1*max(distmatrix))
```

Arguments

`distmatrix`	symmetric (non-negative) distance matrix.
`maxdist`	non-negative numeric. Larger distances are transformed to constant 1.

Details

Transforms large distances to 1, 0 to 0 and continuously linear between 0 and `maxdist`.

Value

A symmetrical matrix.

Author(s)

Christian Hennig <christian.hennig@unibo.it> https://www.unibo.it/sitoweb/christian.hennig/en

See Also

geco

Examples

```
options(digits=4)
data(waterdist)
piecewiselin(waterdist)
```

plotdistreg *Plots for within-groups and between-groups distance regression*

Description

Visualisation of various regressions on distance (or dissimilarity) data where objects are from two groups.

Usage

```
plotdistreg(dmx,dmy,grouping,groups=levels(as.factor(grouping))[1:2],
                        cols=c(1,2,3,4),
                        pchs=rep(1,3),
                        ltys=c(1,2,1,2),
                        individual=TRUE,jointwithin=TRUE,jointall=TRUE,
                        oneplusjoint=TRUE,jittering=TRUE,bcenterline=TRUE,
                        xlim=NULL,ylim=NULL,xlab="geographical distance",
                        ylab="genetic distance",...)
```

Arguments

dmx	dissimilarity matrix or object of class dist. Explanatory dissimilarities (often these will be proper distances, but more general dissimilarities that do not necessarily fulfill the triangle inequality can be used, same for dmy).
dmy	dissimilarity matrix or object of class dist. Response dissimilarities.
grouping	something that can be coerced into a factor, defining the grouping of objects represented by the dissimilarities dmx and dmy (i.e., if grouping has length n, dmx and dmy must be dissimilarities between n objects).
groups	Vector of two levels. The two groups defining the regressions to be compared in the test. These can be factor levels, integer numbers, or strings, depending on the entries of grouping.
cols	vector of four colors (or color numbers) to be used for plotting distances and regression lines within the first group, within the second group, distances between groups, and a line marking the center of the between-groups explanatory distances, see col-argument of par.
pchs	vector of three plot symbols (or numbers) to be used for plotting distances within the first group, within the second group, and distances between groups, see pch-argument of par.
ltys	vector of line type numbers to be used for single group within-group regression, both groups combined within-group regression, regression with all distances, and regression combining within-groups distances of one group with between-groups distances, see lty-argument of par.
individual	if TRUE, within-groups distances regression lines are shown for both groups.
jointwithin	if TRUE, the within-groups distances regression line for both groups combined is shown.

jointall	if TRUE, the regression line based on all distances is shown.
oneplusjoint	if TRUE, the regression lines combining within-groups distances of one group with between-groups distances are shown (the colors of these are the colors of the individual groups, the first two components of the cols-argument).
jittering	if TRUE, points are jittered to avoid overplotting.
bcenterline	if TRUE, a line is plotted to mark the center of the between-groups distances on the explanatory variable.
xlim	to be passed on to plot; the default is determined from the involved distances.
ylim	to be passed on to plot; the default is determined from the involved distances.
xlab	to be passed on to plot.
ylab	to be passed on to plot.
...	optional arguments to be passed on to plot.

Author(s)

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References

Hausdorf, B. and Hennig, C. (2019) Species delimitation and geography. Submitted.

See Also

regeqdist, regdistbetween, regdistbetweenone, regdistdiffone

Examples

```
options(digits=4)
data(veronica)
ver.geo <- coord2dist(coordmatrix=veronica.coord[173:207,],file.format="decimal2")
vei <- prabinit(prabmatrix=veronica[173:207,],distance="jaccard")

species <-c(rep(1,13),rep(2,22))
loggeo <- log(ver.geo+quantile(as.vector(as.dist(ver.geo)),0.25))
plotdistreg(dmx=loggeo,dmy=vei$distmat,grouping=species,
jointwithin=FALSE,jointall=FALSE,groups=c(1,2))
legend(5,0.75,c("within species 1",
"within species 2","species 1 and between","species 2 and between"),lty=c(1,1,2,2),col=c(1,2,1,2))
plotdistreg(dmx=loggeo,dmy=vei$distmat,grouping=species,
jointwithin=TRUE,jointall=TRUE,oneplusjoint=FALSE,groups=c(1,2))
legend(5,0.75,c("within species 1",
"within species 2","all distances","all within species"),lty=c(1,1,1,2),col=c(1,2,3,3))
```

pop.sim *p-value simulation for presence-absence matrices clustering test*

Description

Parametric bootstrap simulation of the p-value of a test of a homogeneity hypothesis against clustering (or significant nestedness). Designed for use within prabtest. The null model is defined by randpop.nb.

Usage

```
pop.sim(regmat, neighbors, h0c = 1, times = 200, dist = "kulczynski",
teststat = "isovertice", testc = NULL, geodist=NULL, gtf=0.1,
n.species = ncol(regmat),
specperreg = NULL, regperspec = NULL, species.fixed=FALSE, pdfnb=FALSE,
ignore.richness=FALSE)
```

Arguments

regmat	0-1-matrix. Columns are species, rows are regions.
neighbors	A list with a component for every region. The components are vectors of integers indicating neighboring regions. A region without neighbors (e.g., an island) should be assigned a list numeric(0).
h0c	numerical. Parameter p.nb for use in randpop.nb.
times	integer. Number of simulation runs.
dist	"kulczynski", "jaccard" or "geco", see kulczynski, geco and jaccard.
teststat	"isovertice", "lcomponent", "distratio", "nn" or "inclusions". See the corresponding functions, homogen.test for "isovertice", incmatrix for "inclusions").
testc	numerical. Tuning constant for the test statistics.
geodist	matrix of non-negative reals. Geographical distances between regions. Only used if dist="geco".
gtf	tuning constant for geco-distance if dist="geco", see "geco".
n.species	integer. Number of species.
specperreg	vector of integers. Numbers of species per region (is calculated from the data by default).
regperspec	vector of integers. Number of regions per species (is calculated from the data by default).
species.fixed	logical. If TRUE, the sizes of the species are taken directly from regmat. Otherwise, they are drawn by random from the empirical distribution of the values from regmat.
pdfnb	logical. Probability correction in randpop.nb.
ignore.richness	logical. If TRUE, there is no assumption of species richnesses to differ between regions in the null model. Regionwise probabilities don't differ in the generation of null data.

Value

List with components

results	vector of teststatistic values for the simulated matrices.
p.above	p-value if large test statistic leads to rejection.
p.below	p-value if small test statistic leads to rejection.
datac	test statistic value for the original data.
testc	see above.

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References

Hennig, C. and Hausdorf, B. (2004) Distance-based parametric bootstrap tests for clustering of species ranges. *Computational Statistics and Data Analysis* 45, 875-896. http://stat.ethz.ch/Research-Reports/110.html.

Hausdorf, B. and Hennig, C. (2003) Biotic Element Analysis in Biogeography. *Systematic Biology* 52, 717-723.

Hausdorf, B. and Hennig, C. (2003) Nestedness of north-west European land snail ranges as a consequence of differential immigration from Pleistocene glacial refuges. *Oecologia* 135, 102-109.

See Also

prabtest, randpop.nb, jaccard, kulczynski, homogen.test, lcomponent, distratio, nn, incmatrix.

Examples

```
options(digits=4)
data(kykladspecreg)
data(nb)
set.seed(1234)
pop.sim(t(kykladspecreg), nb, times=5, h0c=0.35, teststat="nn", testc=3)
```

---

prab.sarestimate *Estimates SAR model from log-abundance matrix of prab-object.*

Description

This is either an interface for the function errorsarlm for abundance data stored in an object of class prab implemented for use in abundtest, or, in case that spatial information should be ignored, it estimates a two-way additive unreplicated linear model for log-abundances on factors species and region.

Usage

```
prab.sarestimate(abmat, prab01=NULL,sarmethod="eigen",
                              weightstyle="C",
                              quiet=TRUE, sar=TRUE,
                              add.lmobject=TRUE)
```

Arguments

abmat	object of class prab.
prab01	presence-absence matrix of same dimensions than the abundance matrix of prabobj. This specifies the presences and absences on which the presence/absence step of abundance-based tests is based (see details). If NULL (which is usually the only reasonable choice), prab01 is computed in order to indicate the nonzeroes of prabobj$prab.
sarmethod	this is passed on as parameter method to errorsarlm and documented there. We don't have experience with any other choice than "eigen".
weightstyle	can take values "W", "B", "C", "U", and "S" though tests suggest that "C" should be chosen. See nb2listw.
quiet	this is passed on as parameter quiet to errorsarlm and documented there.
sar	logical. If TRUE, a simultaneous autoregression model is fitted by calling errorsarlm. If FALSE, a two-way additive unreplicated linear model for log-abundances on factors species and region is computed by lm, ignoring the spatial arrangement of the regions.
add.lmobject	logical. If TRUE, the whole output object of errorsarlm (or lm) is given out.

Value

A list with the following components:

sar	see above.
intercept	numeric. Estimator of the intercept.
sigma	numeric. Estimator of error standard deviation.
regeffects	numeric vector. Estimator for region effects.
speceffects	numeric vector. Estimator for species effects.
lamda	numeric. Governs the degree of spatial autocorrelation. See errorsarlm.
size	integer. Length of neighborhood list generated by nb2listw used by errorsarlm.
nbweight	numeric. Average weight of neighbors.
lmobject	if add.lmobject=TRUE, output object of either lm or errorsarlm.

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See Also

errorsarlm, abundtest

Examples

```
  options(digits=4)
  data(siskiyou)
  x <- prabinit(prabmatrix=siskiyou, neighborhood=siskiyou.nb,
             distance="none")
#  Not run; this needs package spdep
#  prab.sarestimate(x)
  prab.sarestimate(x, sar=FALSE)
```

---

prabclust — *Clustering for biotic elements or for species delimitation (mixture method)*

Description

Clusters a presence-absence matrix object (for clustering ranges/finding biotic elements, Hennig and Hausdorf, 2004) or an object of genetic information (for species delimitation, Hausdorf and Hennig, 2010) by calculating an MDS from the distances, and applying maximum likelihood Gaussian mixtures clustering with "noise" (package mclust) to the MDS points. The solution is plotted.
A standard execution (using the default distance of prabinit) will be
prabmatrix <- prabinit(file="path/prabmatrixfile", neighborhood="path/neighborhoodfile")
clust <- prabclust(prabmatrix)
print(clust)
Examples for species delimitation are given below in the examples section. **Note:** Data formats are described on the prabinit and alleleinit help pages. You may also consider the example datasets kykladspecreg.dat, nb.dat, Heterotrigona_indoFO.txt or MartinezOrtega04AFLP.dat.
Note: prabclust calls the function mclustBIC in package mclust. An alternative is the use of hprabclust.

Usage

```
prabclust(prabobj, mdsmethod = "classical", mdsdim = 4, nnk =
ceiling(prabobj$n.species/40), nclus = 0:9, modelid = "all", permutations=0)

## S3 method for class 'prabclust'
print(x, bic=FALSE, ...)
```

Arguments

prabobj	object of class prab as generated by prabinit. Presence-absence data to be analyzed. (This can be geographical information for range clustering Can also be an object of class alleleobject as generated by alleleinit.

mdsmethod — "classical", "kruskal", or "sammon". The MDS method to transform the distances to data points. "classical" indicates metric MDS by function cmdscale, "kruskal" is non-metric MDS.

mdsdim — integer. Dimension of the MDS points. For mdsmethod=="kruskal", stressvals can be used to see how the stress depends on mdsdim in order to choose mdsdim to get a small stress (smaller than 5%, say).

nnk — integer. Number of nearest neighbors to determine the initial noise estimation by NNclean. nnk=0 fits the model without a noise component.

nclus — vector of integers. Numbers of clusters to perform the mixture estimation.

modelid — string. Model name for mclustBIC (see the corresponding help page; all models or combinations of models mentioned there are possible). modelid="all" compares all possible models. Additionally, "noVVV" is possible, which fits all methods except "VVV".

permutations — integer. It has been found occasionally that depending on the order of observations the algorithms isoMDS and mclustBIC converge to different solutions. This is because these methods require an ordering of the distances, which, if equal distance values are involved, may depend on the order. prabclust uses a standard ordering which should give a reproducible solution in these cases as well. However, if permutations>0, which gives a number of random permutations of the observations, the algorithm is carried out for every permutation and the best solution (in terms of the BIC, based on the lowest stress MDS configuration) is given out (for many datasets this won't change anything except increasing the computing time).

x — object of class prabclust. Output of prabclust.

bic — logical. If TRUE, information about the BIC criterion to choose the model is displayed.

... — necessary for summary method.

Details

Note that if mdsmethod!="classical", zero distances between non-identical objects are replaced by the smallest nonzero distance divided by 10 to prevent the MDS methods from producing an error.

Value

print.prabclust does not produce output. prabclust generates an object of class prabclust. This is a list with components

clustering — vector of integers indicating the cluster memberships of the species. Noise can be recognized by output component symbols.

clustsummary — output object of summary.mclustBIC. A list giving the optimal (according to BIC) parameters, conditional probabilities 'z', and loglikelihood, together with the associated classification and its uncertainty. Note that the numbering of clusters may differ from clustering, see csreorder.

bicsummary — output object of mclustBIC. Bayesian Information Criterion for the specified mixture models and numbers of clusters.

points	numerical matrix. MDS configuration.
nnk	see above.
mdsdim	see above.
mdsmethod	see above.
symbols	vector of characters, similar to clustering, but indicating estimated noise and points belonging to one-point-components (which should be interpreted as some kind of noise as well) by "N".
permchange	logical. If TRUE, permutations>0 has been used and the best solution is different from the one obtained by the standard ordering. (This is just for information and has no further operational consequences.)

Note

Note that we used mdsmethod="kruskal" in our publications, but mdsmethod="classical" is now the default, because of occasional numerical instabilities of the isoMDS-implementation for Jaccard, Kulczynski or geco distance matrices.

Sometimes, prabclust produces an error because mclustBIC cannot handle all models properly. In this case we recommend to change the modelid parameter. "noVVV" and "VVV" are reasonable alternative choices (one of these is expected to reproduce the error, but the other one might work).

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References

Fraley, C. and Raftery, A. E. (1998) How many clusters? Which clustering method? - Answers via Model-Based Cluster Analysis. *Computer Journal* 41, 578-588.

Hausdorf, B. and Hennig, C. (2010) Species Delimitation Using Dominant and Codominant Multilocus Markers. *Systematic Biology*, 59, 491-503.

Hennig, C. and Hausdorf, B. (2004) Distance-based parametric bootstrap tests for clustering of species ranges. *Computational Statistics and Data Analysis* 45, 875-896. http://stat.ethz.ch/Research-Reports/110.html.

See Also

mclustBIC, summary.mclustBIC, NNclean, cmdscale, isoMDS, sammon, prabinit, hprabclust, alleleinit, stressvals.

Examples

```
# Biotic element/range clustering:
data(kykladspecreg)
data(nb)
set.seed(1234)
x <- prabinit(prabmatrix=kykladspecreg, neighborhood=nb)
# If you want to use your own ASCII data files, use
```

```
# x <- prabinit(file="path/prabmatrixfile",
# neighborhood="path/neighborhoodfile")
print(prabclust(x))

# Here is an example for species delimitation with codominant markers;
# only 50 individuals were used in order to have a fast example.
data(tetragonula)
ta <- alleleconvert(strmatrix=tetragonula[1:50,])
tai <- alleleinit(allelematrix=ta)
print(prabclust(tai))

# Here is an example for species delimitation with dominant markers;
# only 50 individuals were used in order to have a fast example.
# You may want to use stressvals to choose mdsdim.
data(veronica)
vei <- prabinit(prabmatrix=veronica[1:50,],distance="jaccard")
print(prabclust(vei,mdsmethod="kruskal",mdsdim=3))
```

`prabinit` *Presence-absence/abundance matrix initialization*

Description

`prabinit` converts a matrix into an object of class `prab` (presence-absence). The matrix may be read from a file or an R-object. It may be a 0-1 matrix or a matrix with non-negative entries (usually abundances). `print.prab` is a print method for such objects.

Documentation here is in terms of biotic elements analysis (species are to be clustered). For species delimitation with dominant markers, see Hausdorf and Hennig (2010), individuals take the role of species and loci take the role of regions.

Usage

```
prabinit(file = NULL, prabmatrix = NULL, rows.are.species = TRUE,
neighborhood = "none", nbbetweenregions=TRUE, geodist=NULL, gtf=0.1,
distance = "kulczynski", toprab = FALSE, toprabp
= 0.05, outc = 5.2)

## S3 method for class 'prab'
print(x, ...)
```

Arguments

`file`	string. non-negative matrix ASCII file (such as example dataset `kykladspecreg.dat`) from which the matrix is read by `read.table`. The usual interpretation is that it is a species-by-regions matrix of species presences/absences (0-1 matrix) or abundances.
`prabmatrix`	matrix with non-negative entries. Either `file` or `prabmatrix` should be `NA`.

rows.are.species
: logical. If TRUE, rows are interpreted as species and columns are interpreted as regions. In this case, rows and columns are interchanged by prabinit.

neighborhood
: A string or a list with a component for every region. The components are vectors of integers indicating neighboring regions. A region without neighbors (e.g., an island) should be assigned a vector numeric(0). If neighborhood is a filename, it is attempted to read such a list from a file, where every row should correspond to one region (such as example dataset nb.dat). If neighborhood="none", all neighborhoods are set to numeric(0). The neighborhood can be tested by nbtest for consistency.

nbbetweenregions
: logical. If TRUE, the neighborhood is defined between regions as explained above. Otherwise it is defined between species (or individuals, if this is used for species delimitation).

geodist
: matrix of non-negative reals. Geographical distances between regions. Only used if distance="geco".

gtf
: tuning constant for geco-distance if distance="geco", see geco.

distance
: "kulczynski", "jaccard", "geco", "qkulczynski", "logkulczynski" (this calls function qkulczynski with log.distance=TRUE), "dice", or "none". The distance measure between species to compute by prabinit.

toprab
: logical. If TRUE, a presence-absence matrix is computed from the non-negative input matrix. "Absence", i.e., the entry 0, is chosen if the original entry is 0, or the original entry is smaller than or equal to toprabp times the sum of entries in the corresponding region, and log(original entry) is considered to be a lower outlier compared with the other entries of the corresponding species (see outc). "Presence", i.e., the entry 1, thus means that the original entry is non-negligible w.r.t. the species or w.r.t. the region.

toprabp
: numerical between 0 and 1, see toprab.

outc
: numerical. Tuning constant for the outlier identification associated with toprab=TRUE. An entry smaller than or equal to outc*mad times the median is considered as a lower outlier.

x
: object of class prab.

...
: necessary for print method.

Details

Species that are absent in all regions are omitted.

Value

prabinit produces an object of class prab, which is a list with components

distmat
: distance matrix between species.

prab
: abundance or presence/absence matrix (if presence/absence, the entries are logical). Rows are regions, columns are species.

nb
: neighborhood list, see above.

`regperspec` vector of the number of regions occupied by a species.

`specperreg` vector of the number of species present in a region.

`n.species` number of species (in the `prab`-object, see `nonzero`).

`n.regions` number of regions.

`distance` string denoting the chosen distance measure.

`geodist` non-negative matrix. see above.

`gtf` numeric. see above.

`spatial` `TRUE`, if there is a specified neighborhood structure.

`nonempty.species`
logical vector. The length is the number of species in the original file/matrix. If `FALSE`, the corresponding species had only zero entries and was therefore absent. Note that these species are not included in any other component of a `prab` object, i.e., `n.species` is the number of `TRUE`-entries in `nonzero`.

`nbbetweenregions`
see above.

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References

Hausdorf, B. and Hennig, C. (2010) Species Delimitation Using Dominant and Codominant Multilocus Markers. *Systematic Biology*, 59, 491-503.

See Also

`read.table`, `jaccard`, `kulczynski`, `geco`, `qkulczynski`, `nbtest`, `alleleinit`

Examples

```
# If you want to use your own ASCII data files, use
# x <- prabinit(file="path/prabmatrixfile",
# neighborhood="path/neighborhoodfile")
data(kykladspecreg)
data(nb)
prabinit(prabmatrix=kykladspecreg, neighborhood=nb)
```

prabtest *Parametric bootstrap test for clustering in presence-absence matrices*

Description

Parametric bootstrap test of a null model of i.i.d., but spatially autocorrelated species against clustering of the species' occupied areas (or alternatively nestedness). In spite of the lots of parameters, a standard execution (for the default test statistics, see parameter `teststat` below) will be

```
prabmatrix <- prabinit(file="path/prabmatrixfile", neighborhood="path/neighborhoodfile")
test <- prabtest(prabmatrix)
summary(test)
```

Note: Data formats are described on the `prabinit` help page. You may also consider the example datasets `kykladspecreg.dat` and `nb.dat`. Take care of the parameter `rows.are.species` of `prabinit`.

Usage

```
prabtest(prabobject, teststat = "distratio", tuning = switch(teststat,
    distratio = 0.25, lcomponent = floor(3 * ncol(prabobject$distmat)/4),
    isovertice = ncol(prabobject$distmat), nn = 4, NA), times = 1000,
    pd = NULL, prange = c(0, 1), nperp = 4, step = 0.1, step2=0.01,
                      twostep = TRUE,
    sf.sim = FALSE, sf.const = sf.sim, pdfnb = FALSE, ignore.richness=FALSE)


## S3 method for class 'prabtest'
summary(object, above.p=object$teststat %in%
         c("groups","inclusions","mean"),
         group.outmean=FALSE,...)

## S3 method for class 'summary.prabtest'
print(x, ...)
```

Arguments

`prabobject`	an object of class `prab` (presence-absence data), as generated by `prabinit`.
`teststat`	string, indicating the test statistics. `"isovertice"`: number of isolated vertices in the graph of `tuning` smallest distances between species. `"lcomponent"`: size of largest connectivity component in this graph. `"distratio"`: ratio between `tuning` smallest and largest distances. `"nn"`: average distance of species to `tuning`th nearest neighbor. `"inclusions"`: number of inclusions between areas of different species (tests for nestedness structure, not for clustering).
`tuning`	integer or (if `teststat="distratio"`) numerical between 0 and 1. Tuning constant for test statistics, see `teststat`.
`times`	integer. Number of simulation runs.

pd	numerical between 0 and 1. The probability that a new region is drawn from the non-neighborhood of the previous regions belonging to a species under generation. If NA (the default), prabtest estimates this by function autoconst. Otherwise the next five parameters have no effect.
prange	numerical range vector, lower value not smaller than 0, larger value not larger than 1. Range where pd is to be found. Used by function autoconst.
nperp	integer. Number of simulations per pd-value. Used by function autoconst.
step	numerical between 0 and 1. Interval length between subsequent choices of pd for the first simulation. Used by function autoconst.
step2	numerical between 0 and 1. Interval length between subsequent choices of pd for the second simulation (see parameter twostep). Used by function autoconst.
twostep	logical. If TRUE, a first estimation step for pd is carried out in the whole prange, and then the final estimation is determined between the preliminary estimator -5*step2 and +5*step2. Else, the first simulation determines the final estimator. Used by function autoconst.
sf.sim	logical. Indicates if the range sizes of the species are held fixed in the test simulation (TRUE) or generated from their empirical distribution in x (FALSE). See function randpop.nb.
sf.const	logical. Same as sf.sim, but for estimation of pd by autoconst.
pdfnb	logical. If TRUE, the probabilities of the regions are modified according to the number of neighboring regions in randpop.nb, see Hennig and Hausdorf (2002), p. 5. This is usually no improvement.
ignore.richness	logical. If TRUE, there is no assumption of species richnesses to differ between regions in the null model. Regionwise probabilities don't differ in the generation of null data.
object	object of class prabtest.
above.p	logical. TRUE means that for output from abundtest the p-value is p.above, otherwise p.below.
group.outmean	logical. If TRUE and object$teststat="groups", statistics concerning the mean of all dissimilarities are given out by print.summary.prabtest.
x	object of class summary.prabtest.
...	no meaning, necessary for print and summary methods.

Details

From the original data, the distribution of the range sizes of the species, the autocorrelation parameter pd (estimated by autoconst) and the distribution on the regions induced by the relative species numbers are taken. With these parameters, times populations according to the null model implemented in randpop.nb are generated and the test statistic is evaluated. The resulting p-value is number of simulated statistic values more extreme than than the value of the original data+1 divided by times+1. "More extreme" means smaller for "lcomponent", "distratio", "nn", larger for "inclusions", and twice the smaller number between the original statistic value and the "border", i.e., a two-sided test for "isovertice". If pd=NA was specified, a diagnostic plot for the estimation of pd is plotted by autoconst. For details see Hennig and Hausdorf (2004) and the help pages of the cited functions.

Value

prabtest prodices an object of class prabtest, which is a list with components

results	vector of test statistic values for all simulated populations.
datac	test statistic value for the original data.'
p.value	the p-value.
tuning	see above.
pd	see above.
reg	regression coefficients from autoconst.
teststat	see above.
distance	the distance measure chosen, see prabinit.
gtf	the geco-distance tuning parameter (only informative if distance="geco"), see prabinit.
times	see above.
pdfnb	see above.
ignore.richness	see above.

summary.prabtest produces an object of class summary.prabtest, which is a list with components

rrange	range of the simulation results (test statistic values) of object.
rmean	mean of the simulation results (test statistic values) of object.
datac, p.value, pd, tuning, teststat, distance, times, pdfnb, abund, sarlambda	directly taken from object, see prabtest and abundtest.
groupinfo	if object$teststat="groups", components rrangeg (matrix of group-wise ranges of test statistic value), rmeang (vector of group-wise means of test statistic value), rrangem (range over simulations of overall mean of within-group dissimilarities), rmeanm (mean over simulations of overall mean of within-group dissimilarities) are added to the list object$groupinfo, and this is given out.

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References

Hennig, C. and Hausdorf, B. (2004) Distance-based parametric bootstrap tests for clustering of species ranges. *Computational Statistics and Data Analysis* 45, 875-896. http://stat.ethz.ch/Research-Reports/110.html.

Hausdorf, B. and Hennig, C. (2003) Biotic Element Analysis in Biogeography. *Systematic Biology* 52, 717-723.

Hausdorf, B. and Hennig, C. (2003) Nestedness of north-west European land snail ranges as a consequence of differential immigration from Pleistocene glacial refuges. *Oecologia* 135, 102-109.

See Also

prabinit generates objects of class prab.

autoconst estimates pd from such objects.

randpop.nb generates populations from the null model. An alternative model is given by cluspop.nb.

Some more information on the test statistics is given in homogen.test, lcomponent, distratio, nn, incmatrix.

The simulations are computed by pop.sim.

Examples

```
options(digits=4)
data(kykladspecreg)
data(nb)
set.seed(1234)
x <- prabinit(prabmatrix=kykladspecreg, neighborhood=nb)
# If you want to use your own ASCII data files, use
# x <- prabinit(file="path/prabmatrixfile",
# neighborhood="path/neighborhoodfile")
kpt <- prabtest(x, times=5, pd=0.35)
# These settings are chosen to make the example execution
# a bit faster; usually you will use prabtest(kprab).
summary(kpt)
```

---

qkulczynski *Quantitative Kulczynski distance matrix*

Description

Computes quantitative Kulczynski distances between the columns of an abundance matrix.

Usage

```
qkulczynski(regmat, log.distance=FALSE)
```

Arguments

regmat	(non-negative) abundance matrix. Columns are species, rows are regions.
log.distance	logical. If TRUE, 1 is added to the abundance matrix and then the logs of the values are taken in order to compute the distance.

Details

The quantitative Kulczynski distance between two species is 1-(mean of (mean of over regions minimum abundance of both species)/(sum of abundances of species 1) and (mean of over regions minimum abundance of both species)/(sum of abundances of species 2)). If the abundance matrix is a 0-1-matrix, this gives the standard Kulczynski distance.

Value

A symmetrical matrix of quantitative Kulczynski distances.

Author(s)

Christian Hennig <christian.hennig@unibo.it> https://www.unibo.it/sitoweb/christian.hennig/en

References

D. P. Faith, P. R. Minchin and L. Belbin (1987) Compositional dissimilarity as a robust measure of ecological distance. *Vegetation* 69, 57-68.

See Also

kulczynski

Examples

```
options(digits=4)
data(kykladspecreg)
qkulczynski(t(kykladspecreg))
```

---

randpop.nb *Simulation of presence-absence matrices (non-clustered)*

Description

Generates a simulated matrix where the rows are interpreted as regions and the columns as species, 1 means that a species is present in the region and 0 means that the species is absent. Species are generated i.i.d.. Spatial autocorrelation of a species' presences is governed by the parameter p.nb and a list of neighbors for each region.

Usage

```
randpop.nb(neighbors, p.nb = 0.5, n.species, n.regions =
length(neighbors), vector.species = rep(1, n.species),
species.fixed = FALSE, pdf.regions = rep(1/n.regions, n.regions),
count = TRUE, pdfnb = FALSE)
```

Arguments

neighbors	A list with a component for every region. The components are vectors of integers indicating neighboring regions. A region without neighbors (e.g., an island) should be assigned a list numeric(0).

p.nb	numerical between 0 and 1. The probability that a new region is drawn from the non-neighborhood of the previous regions belonging to a species under generation. Note that for a given presence-absence matrix, this parameter can be estimated by autoconst (called pd there).
n.species	integer. Number of species.
n.regions	integer. Number of regions.
vector.species	vector of integers. If species.fixed=TRUE, vector.species must have length n.species and gives the sizes (i.e., numbers of regions) of the species to generate. Else, the sizes are generated randomly from the empirical distribution of vector.species.
species.fixed	logical. See vector.species.
pdf.regions	numerical vector of length n.species. The entries must sum up to 1 and give probabilities for the regions to be drawn during the generation of a species. These probabilities are used conditional on the new region being a neighbor or a non-neighbor of the previous regions of the species, see p.nb.
count	logical. If TRUE, the number of the currently generated species is printed.
pdfnb	logical. If TRUE, the probabilities of the regions are modified according to the number of neighboring regions by dividing them relative to the others by min(1,number of neighbors).

Details

The principle is that a single species with given size is generated one-by-one region. The first region is drawn according to pdf.regions. For all following regions, a neighbor or non-neighbor of the previous configuration is added (if possible), as explained in pdf.regions, p.nb.

Value

A 0-1-matrix, rows are regions, columns are species.

Author(s)

Christian Hennig <christian.hennig@unibo.it> https://www.unibo.it/sitoweb/christian.hennig/en

References

Hennig, C. and Hausdorf, B. (2004) Distance-based parametric bootstrap tests for clustering of species ranges. *Computational Statistics and Data Analysis* 45, 875-896. http://stat.ethz.ch/Research-Reports/110.html.

Hausdorf, B. and Hennig, C. (2003) Biotic Element Analysis in Biogeography. *Systematic Biology* 52, 717-723.

Hausdorf, B. and Hennig, C. (2003) Nestedness of nerth-west European land snail ranges as a consequence of differential immigration from Pleistocene glacial refuges. *Oecologia* 135, 102-109.

See Also

autoconst estimates p.nb from matrices of class prab. These are generated by prabinit.

prabtest uses randpop.nb as a null model for tests of clustering. An alternative model is given by cluspop.nb.

Examples

```
data(nb)
set.seed(2346)
randpop.nb(nb, p.nb=0.1, n.species=5, vector.species=c(1,10,20,30,34))
```

---

regdist *Regression between subsets of dissimilarity matrices*

Description

Given two dissimilarity matrices dmx and dmy and an indicator vector x, this computes a standard least squares regression between the dissimilarity between objects indicated in x.

Usage

```
regdist(x,dmx,dmy,xcenter=0,param)
```

Arguments

x	vector of logicals of length of the number of objects on which dissimilarities dmx and dmy are based.
dmx	dissimilarity matrix or object of class dist. Explanatory dissimilarities.
dmy	dissimilarity matrix or object of class dist. Response dissimilarities.
xcenter	numeric. Dissimilarities dmx are centered by this, i.e., this value is subtracted from the dissimilarities before regression.
param	1 or 2 or NULL. If 1 or 2, only the first or second parameter (intercept or slope) of the regression is given out.

Value

If param=NULL, the output object of lm. If param=1 the intercept. If param=2 the slope.

Author(s)

Christian Hennig <christian.hennig@unibo.it> https://www.unibo.it/sitoweb/christian.hennig/en

References

Hausdorf, B. and Hennig, C. (2019) Species delimitation and geography. Submitted.

Examples

```
options(digits=4)
data(veronica)
ver.geo <- coord2dist(coordmatrix=veronica.coord[1:20,],file.format="decimal2")
vei <- prabinit(prabmatrix=veronica[1:20,],distance="jaccard")
regdist(c(rep(TRUE,10),rep(FALSE,10)),ver.geo,vei$distmat,param=1)
```

`regdistbetween` *Testing equality of within-groups and between-groups distances regression*

Description

Jackknife-based test for equality of two regressions between distances. Given two groups of objects, this tests whether the regression involving all distances is compatible with the regression involving within-group distances only.

Usage

```
regdistbetween(dmx,dmy,grouping,groups=levels(as.factor(grouping))[1:2])

## S3 method for class 'regdistbetween'
print(x,...)
```

Arguments

`dmx`	dissimilarity matrix or object of class `dist`. Explanatory dissimilarities (often these will be proper distances, but more general dissimilarities that do not necessarily fulfill the triangle inequality can be used, same for `dmy`).
`dmy`	dissimilarity matrix or object of class `dist`. Response dissimilarities.
`grouping`	something that can be coerced into a factor, defining the grouping of objects represented by the dissimilarities `dmx` and `dmy` (i.e., if `grouping` has length n, `dmx` and `dmy` must be dissimilarities between `n` objects).
`groups`	Vector of two levels. The two groups defining the regressions to be compared in the test. These can be factor levels, integer numbers, or strings, depending on the entries of `grouping`.
`x`	object of class `"regdistbetween"`.
`...`	optional arguments for print method.

Details

The null hypothesis that the regressions based on all distances and based on within-group distances only are equal is tested using jackknife pseudovalues. This assumes that a single regression is appropriate at least for the within-group distances alone. The test statistic is the difference between fitted values with x (explanatory variable) fixed at the center of the between-group distances. The

test is run one-sided, i.e., the null hypothesis is only rejected if the between-group distances are larger than expected under the null hypothesis, see below.

The test cannot be run in case that within-group regressions or jackknifed within-group regressions are ill-conditioned.

This was implemented having in mind an application in which the explanatory distances represent geographical distances, the response distances are genetic distances, and groups represent species or species-candidates. In this application, for testing whether the regression patterns are compatble with the two groups behaving like a single species, one would first use `regeqdist` to test whether a joint regression for the within-group distances of both groups makes sense. If this is not rejected, `regdistbetween` is run to see whether the between-group distances are compatible with the within-group distances. This is only rejected if the between-group distances are larger than expected under equality of regressions, because if they are smaller, this is not an indication against the groups belonging together genetically.

If a joint regression on within-group distances is rejected by `regeqdist`, `regdistbetweenone` can be used to test whether the between-group distances are at least compatible with the within-group distances of one of the groups, which can still be the case within a single species, see Hausdorf and Hennig (2019).

Value

list of class `"regdistbetween"` with components

`pval`	p-value.
`coeffdiff`	difference between regression fits (all distances minus within-group distances only) at `xcenterbetween`, see below.
`condition`	condition numbers of regressions, see `kappa`.
`lmfit`	list. Output objects of `lm` within the two groups.
`jr`	output object of `jackknife` for difference between regression fitted values at `xcenterbetween`.
`xcenter`	mean of within-groups distances of explanatory variable, used for centering.
`xcenterbetween`	mean of between-groups distances of explanatory variable (after centering by `xcenter`); at this point regression fitted values are computed.
`tstat`	t-statistic.
`tdf`	degrees of freedom of t-statistic.
`jackest`	jackknife-estimator of difference between regression fitted values at `xcenterbetween`.
`jackse`	jackknife-standard error for `jackest`.
`jackpseudo`	vector of jacknife pseudovalues on which the test is based.
`testname`	title to be printed out when using `print.regdistbetween`.
`groups`	see above.

Author(s)

Christian Hennig <christian.hennig@unibo.it> https://www.unibo.it/sitoweb/christian.hennig/en

References

Hausdorf, B. and Hennig, C. (2019) Species delimitation and geography. Submitted.

See Also

regeqdist, regdistbetweenone

Examples

```
options(digits=4)
data(veronica)
ver.geo <- coord2dist(coordmatrix=veronica.coord[173:207,],file.format="decimal2")
vei <- prabinit(prabmatrix=veronica[173:207,],distance="jaccard")
loggeo <- log(ver.geo+quantile(as.vector(as.dist(ver.geo)),0.25))

species <-c(rep(1,13),rep(2,22))

rtest2 <-
regdistbetween(dmx=loggeo,dmy=vei$distmat,grouping=species,groups=c(1,2))
print(rtest2)
```

---

regdistbetweenone *Testing equality of one within-group and between-two groups distances regression*

Description

Jackknife-based test for equality of two regressions between distances. Given two groups of objects, this tests whether the regression involving the distances within one of the groups is compatible with the regression involving the same within-group distances together with the between group distances.

Usage

```
regdistbetweenone(dmx,dmy,grouping,groups=levels(as.factor(grouping))[1:2],rgroup)
```

Arguments

dmx	dissimilarity matrix or object of class dist. Explanatory dissimilarities (often these will be proper distances, but more general dissimilarities that do not necessarily fulfill the triangle inequality can be used, same for dmy).
dmy	dissimilarity matrix or object of class dist. Response dissimilarities.
grouping	something that can be coerced into a factor, defining the grouping of objects represented by the dissimilarities dmx and dmy (i.e., if grouping has length n, dmx and dmy must be dissimilarities between n objects).
groups	vector of two levels. The two groups defining the regressions to be compared in the test. These can be factor levels, integer numbers, or strings, depending on the entries of grouping.
rgroup	one of the levels in groups, denoting the group of which within-group dissimilarities are considered.

Details

The null hypothesis that the regressions based on the distances within group species and based on these distances together with the between-groups distances are equal is tested using jackknife pseudovalues. The test statistic is the difference between fitted values with x (explanatory variable) fixed at the center of the between-group distances. The test is run one-sided, i.e., the null hypothesis is only rejected if the between-group distances are larger than expected under the null hypothesis, see below. For the jackknife, observations from both groups are left out one at a time. However, the roles of the two groups are different (observations from group species are used in both regressions whereas observations from the other group are only used in one of them), and therefore the corresponding jackknife pseudovalues can have different variances. To take this into account, variances are pooled, and the degrees of freedom of the t-test are computed by the Welch-Sattertwaithe approximation for aggregation of different variances.

The test cannot be run and many components will be NA in case that within-group regressions or jackknifed within-group regressions are ill-conditioned.

This was implemented having in mind an application in which the explanatory distances represent geographical distances, the response distances are genetic distances, and groups represent species or species-candidates. In this application, for testing whether the regression patterns are compatble with the two groups behaving like a single species, one would first use regeqdist to test whether a joint regression for the within-group distances of both groups makes sense. If this is not rejected, regdistbetween is run to see whether the between-group distances are compatible with the within-group distances. If a joint regression on within-group distances is rejected by regeqdist, regdistbetweenone can be used to test whether the between-group distances are at least compatible with the within-group distances of one of the groups, which can still be the case within a single species, see Hausdorf and Hennig (2019). This is only rejected if the between-group distances are larger than expected under equality of regressions, because if they are smaller, this is not an indication against the groups belonging together genetically. To this end, regdistbetweenone needs to be run twice using both groups as species. This will produce two p-values. The null hypothesis that the regressions are compatible for at least one group can be rejected if the maximum of the two p-values is smaller than the chosen significance level.

Value

list of class "regdistbetween" with components

pval	p-value.
coeffdiff	difference between regression fits (within-group together with between-groups distances minus within-group distances only) at xcenterbetween, see below.
condition	condition numbers of regressions, see kappa.
lmfit	list. Output objects of lm within the two groups.
jr	output object of jackknife for difference between regression fitted values at xcenterbetween.
xcenter	mean of within-group distances for group species of explanatory variable, used for centering.
xcenterbetween	mean of between-groups distances of explanatory variable (after centering by xcenter); at this point regression fitted values are computed.
tstat	t-statistic.

tdf	degrees of freedom of t-statistic according to Welch-Sattertwaithe approximation.
jackest	jackknife-estimator of difference between regression fitted values at xcenterbetween.
jackse	jackknife-standard error for jackest.
jackpseudo	vector of jacknife pseudovalues on which the test is based.
groups	see above.
species	see above.
testname	title to be printed out when using print.regdistbetween.

Author(s)

Christian Hennig <christian.hennig@unibo.it> https://www.unibo.it/sitoweb/christian.hennig/en

References

Hausdorf, B. and Hennig, C. (2019) Species delimitation and geography. Submitted.

See Also

regeqdist, regdistbetweenone

Examples

```
options(digits=4)
data(veronica)
ver.geo <- coord2dist(coordmatrix=veronica.coord[173:207,],file.format="decimal2")
vei <- prabinit(prabmatrix=veronica[173:207,],distance="jaccard")

species <-c(rep(1,13),rep(2,22))
loggeo <- log(ver.geo+quantile(as.vector(as.dist(ver.geo)),0.25))
rtest3 <-
regdistbetweenone(dmx=loggeo,dmy=vei$distmat,grouping=species,groups=c(1,2),rgroup=1)
print(rtest3)
```

---

regdistdiff *Regression difference between within-group dissimilarities*

Description

Given two dissimilarity matrices dmx and dmy, an indicator vector x and a grouping, this computes the difference between standard least squares regression predictions at point xcenterbetween. The regressions are based on the dissimilarities in dmx vs. dmy for objects indicated in x. grouping indicates the two groups, and the difference is computed between regressions based on the within-group distances of the two groups.

Usage

```
regdistdiff(x,dmx,dmy,grouping,xcenter=0,xcenterbetween=0)
```

Arguments

x	vector of logicals of length of the number of objects on which dissimilarities dmx and dmy are based.
dmx	dissimilarity matrix or object of class dist. Explanatory dissimilarities.
dmy	dissimilarity matrix or object of class dist. Response dissimilarities.
grouping	vector of length of the number of objects on which dissimilarities dmx and dmy are based. Grouping vector. Regressions will be based on the first two values that appear in unique(grouping[x]) (note that objects that are not assigned to one of these groups will be ignored); normally grouping should indicate only two groups on the objects with x=TRUE, and then these are used.
xcenter	numeric. Dissimilarities dmx are centered by this, i.e., this value is subtracted from the dissimilarities before regression.
xcenterbetween	numeric. This specifies the x- (dissimilarity) value at which predictions from the two regressions are compared. Note that this is interpreted as after centering by xcenter.

Value

Difference between standard least squares regression predictions for the two groups at point xcenterbetween.

Author(s)

Christian Hennig <christian.hennig@unibo.it> https://www.unibo.it/sitoweb/christian.hennig/en

References

Hausdorf, B. and Hennig, C. (2019) Species delimitation and geography. Submitted.

See Also

regdistbetween

Examples

```
options(digits=4)
data(veronica)
ver.geo <- coord2dist(coordmatrix=veronica.coord[173:207,],file.format="decimal2")
vei <- prabinit(prabmatrix=veronica[173:207,],distance="jaccard")

species <-c(rep(1,13),rep(2,22))
regdistdiff(rep(TRUE,35),ver.geo,vei$distmat,grouping=species,xcenter=0,xcenterbetween=100)
```

regdistdiffone *Regression difference within reference group and between-group dissimilarities*

Description

Given two dissimilarity matrices dmx and dmy, an indicator vector x and a grouping, this computes the difference between standard least squares regression predictions at point xcenterbetween. The regressions are based on the dissimilarities in dmx vs. dmy for objects indicated in x. grouping indicates the two groups, and the difference is computed between regressions based on (a) the within-group distances of the reference group rgroup and (b) these together with the between-group distances.

Usage

```
regdistdiffone(x,dmx,dmy,grouping,xcenter=0,xcenterbetween=0,rgroup)
```

Arguments

x	vector of logicals of length of the number of objects on which dissimilarities dmx and dmy are based.
dmx	dissimilarity matrix or object of class dist. Explanatory dissimilarities.
dmy	dissimilarity matrix or object of class dist. Response dissimilarities.
grouping	vector of length of the number of objects on which dissimilarities dmx and dmy are based. Grouping vector. Regressions will be based on the first two values that appear in unique(grouping[x]) (note that objects that are not assigned to one of these groups will be ignored); normally grouping should indicate only two groups on the objects with x=TRUE, and then these are used.
xcenter	numeric. Dissimilarities dmx are centered by this, i.e., this value is subtracted from the dissimilarities before regression.
xcenterbetween	numeric. This specifies the x- (dissimilarity) value at which predictions from the two regressions are compared. Note that this is interpreted as after centering by xcenter.
rgroup	one of the values of grouping, specifying the reference group.

Value

Difference between standard least squares regression predictions for the two regressions at point xcenterbetween.

Author(s)

Christian Hennig <christian.hennig@unibo.it> https://www.unibo.it/sitoweb/christian.hennig/en

References

Hausdorf, B. and Hennig, C. (2019) Species delimitation and geography. Submitted.

See Also

regdistbetweenone

Examples

```
options(digits=4)
data(veronica)
ver.geo <- coord2dist(coordmatrix=veronica.coord[173:207,],
  file.format="decimal2")
vei <- prabinit(prabmatrix=veronica[173:207,],distance="jaccard")

species <-c(rep(1,13),rep(2,22))
regdistdiffone(rep(TRUE,35),ver.geo,vei$distmat,grouping=species,
  xcenter=0,xcenterbetween=100,rgroup=2)
```

---

regeqdist *Testing equality of two distance-regressions*

Description

Jackknife-based test for equality of two regressions between distance matrices.

Usage

```
regeqdist(dmx,dmy,grouping,groups=levels(as.factor(grouping))[1:2])

## S3 method for class 'regeqdist'
print(x,...)
```

Arguments

dmx	dissimilarity matrix or object of class dist. Explanatory dissimilarities (often these will be proper distances, but more general dissimilarities that do not necessarily fulfill the triangle inequality can be used, same for dmy).
dmy	dissimilarity matrix or object of class dist. Response dissimilarities.
grouping	something that can be coerced into a factor, defining the grouping of objects represented by the dissimilarities dmx and dmy (i.e., if grouping has length n, dmx and dmy must be dissimilarities between n objects).
groups	Vector of two, indicating the two groups defining the regressions to be compared in the test. These can be factor levels, integer numbers, or strings, depending on the entries of grouping.
x	object of class "regeqdist".
...	optional arguments for print method.

Details

The null hypothesis that the regressions within the two groups are equal is tested using jackknife pseudovalues independently in both groups allowing for potentially different variances of the pseudovalues, and aggregating as in Welch's t-test. Tests are run separately for intercept and slope and aggregated by Bonferroni's rule.

The test cannot be run and many components will be NA in case that within-group regressions or jackknifed within-group regressions are ill-conditioned.

This was implemented having in mind an application in which the explanatory distances represent geographical distances, the response distances are genetic distances, and groups represent species or species-candidates. In this application, for testing whether the regression patterns are compatble with the two groups behaving like a single species, one would first use `regeqdist` to test whether a joint regression for the within-group distances of both groups makes sense. If this is not rejected, `regdistbetween` is run to see whether the between-group distances are compatible with the within-group distances. On the other hand, if a joint regression on within-group distances is rejected, `regdistbetweenone` can be used to test whether the between-group distances are at least compatible with the within-group distances of one of the groups, which can still be the case within a single species, see Hausdorf and Hennig (2019).

Value

list of class `"regeqdist"` with components

`pval`	p-values for intercept and slope.
`coeffdiff`	vector of differences between groups (first minus second) for intercept and slope.
`condition`	condition numbers of regressions, see `kappa`.
`lmfit`	list. Output objects of `lm` within the two groups.
`jr`	list of two lists of two; output object of `jackknife` within the two groups for intercept and slope.
`xcenter`	mean of `dmx` within the two groups used for centering.
`tstat`	t-statistic.
`tdf`	vector of degrees of freedom of t-statistic according to Welch-Sattertwaithe approximation for intercept and slope.
`jackest`	jackknife-estimator of difference between regressions; vector with intercept and slope difference.
`jackse`	vector with jackknife-standard errors for `jackest`, intercept and slope.
`jackpseudo`	list of two lists of vectors; jacknife pseudovalues within both groups for intercept and slope estimators.
`groups`	see above.

Author(s)

Christian Hennig <christian.hennig@unibo.it> https://www.unibo.it/sitoweb/christian.hennig/en

References

Hausdorf, B. and Hennig, C. (2019) Species delimitation and geography. Submitted.

See Also

regdistbetween, regdistbetweenone

Examples

```
options(digits=4)
data(veronica)
ver.geo <- coord2dist(coordmatrix=veronica.coord[173:207,],file.format="decimal2")
vei <- prabinit(prabmatrix=veronica[173:207,],distance="jaccard")
loggeo <- log(ver.geo+quantile(as.vector(as.dist(ver.geo)),0.25))

species <-c(rep(1,13),rep(2,22))
rtest <- regeqdist(dmx=loggeo,dmy=vei$distmat,grouping=species,groups=c(1,2))
print(rtest)
```

---

regpop.sar *Simulation of abundance matrices (non-clustered)*

Description

Generates a simulated matrix where the rows are interpreted as regions and the columns as species, and the entries are abundances. Species are generated i.i.d. in two steps. In the first step, a presence-absence matrix is generated as in randpop.nb. In the second step, conditionally on presence in the first step, abundance values are generated according to a simultaneous autoregression (SAR) model for the log-abundances (see errorsarlm for the model; estimates are provided by the parameter sarestimate). Spatial autocorrelation of a species' presences is governed by the parameter p.nb, sarestimate and a list of neighbors for each region.

Usage

```
regpop.sar(abmat, prab01=NULL, sarestimate=prab.sarestimate(abmat),
                    p.nb=NULL,
                    vector.species=prab01$regperspec,
                    pdf.regions=prab01$specperreg/(sum(prab01$specperreg)),
                   count=FALSE)
```

Arguments

abmat	object of class prab, containing the abundance or presence/absence data.
prab01	presence-absence matrix of same dimensions than the abundance matrix of prabobj. This specifies the presences and absences on which the presence/absence step of abundance-based tests is based (see details). If NULL (which is usually the only reasonable choice), prab01 is computed in order to indicate the nonzeroes of prabobj$prab.

sarestimate	Estimator of the parameters of a simultaneous autoregression model corresponding to the null model for abundance data from Hausdorf and Hennig (2007) as generated by prab.sarestimate. This requires package spdep. If sarestimate$sar=FALSE, spatial structure is ignored for generating the abundance values.
p.nb	numeric between 0 and 1. The probability that a new region is drawn from the non-neighborhood of the previous regions belonging to a species under generation. If NULL, the spatial structure of the regions is ignored. Note that for a given presence-absence matrix, this parameter can be estimated by autoconst (called pd there).
vector.species	vector of integers. vector.species gives the sizes (i.e., numbers of regions) of the species to generate..
pdf.regions	numerical vector of length n.species. The entries must sum up to 1 and give probabilities for the regions to be drawn during the generation of a species. These probabilities are used conditional on the new region being a neighbor or a non-neighbor of the previous regions of the species, see p.nb.
count	logical. If TRUE, the number of the currently generated species is printed.

Value

A matrix of abundance values, rows are regions, columns are species.

Author(s)

Christian Hennig <christian.hennig@unibo.it> https://www.unibo.it/sitoweb/christian.hennig/en

References

Hausdorf, B. and Hennig, C. (2007) Null model tests of clustering of species, negative co-occurrence patterns and nestedness in meta-communities. *Oikos* 116, 818-828.

See Also

autoconst estimates p.nb from matrices of class prab. These are generated by prabinit.

abundtest uses regpop.sar as a null model for tests of clustering.

randpop.nb (analogous function for simulating presence-absence data)

Examples

```
options(digits=4)
data(siskiyou)
set.seed(1234)
x <- prabinit(prabmatrix=siskiyou, neighborhood=siskiyou.nb,
             distance="none")
# Not run; this needs package spdep.
# regpop.sar(x, p.nb=0.046)
regpop.sar(x, p.nb=0.046, sarestimate=prab.sarestimate(x,sar=FALSE))
```

siskiyou *Herbs of the Siskiyou Mountains*

Description

Distributions of species of herbs in relation to elevation on quartz diorite in the central Siskiyou Mountains. All values are per mille frequencies in transects (The number of 1 m2 quadrats, among 1000 such quadrats, in which a species was observed, based on 1250 1m2 quadrats in the first 5 transects, and 400 1m2 quadrats in 6. transect). Observed presences in the transect, outside the sampling plots, were coded as 0.2. Rows correspond to species, columns correspond to regions.

Usage

```
data(siskiyou)
```

Format

Three objects are generated:

siskiyou numeric matrix giving the 144*6 abundance values.

siskiyou.nb neighborhood list for the 6 regions.

siskiyou.groups integer vector of length 144, giving group memberships for the 144 species.

Details

Reads from example data files `LeiMik1.dat`, `LeiMik1NB.dat`, `LeiMik1G.dat`.

Source

Whittaker, R. H. 1960. Vegetation of the Siskiyou Mountains, Oregon and California. *Ecol. Monogr.* 30: 279-338 (table 14).

Examples

```
data(siskiyou)
```

specgroups *Average within-group distances for given groups*

Description

Generates average within-group distances (overall and group-wise) from a dissimilarity matrix and a given grouping.

Usage

```
specgroups(distmat,groupvector, groupinfo)
```

Arguments

distmat	dissimilarity matrix or dist-object.
groupvector	integer vector. For every row of distmat, a number indicating the group membership.
groupinfo	list with components lg (levels of groupvector), ng (number of groups), nsg (vector of group sizes).

Value

A list with parameters

overall	overall average within-groups dissimilarity.
gr	vector of group-wise average within-group dissimilarities (this will be NaN if the group size is only 1).

Author(s)

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Examples

```
options(digits=4)
data(siskiyou)
x <- prabinit(prabmatrix=siskiyou, neighborhood=siskiyou.nb,
            distance="logkulczynski")
groupvector <- as.factor(siskiyou.groups)
ng <- length(levels(groupvector))
lg <- levels(groupvector)
nsg <- numeric(0)
  for (i in 1:ng) nsg[i] <- sum(groupvector==lg[i])
groupinfo <- list(lg=lg,ng=ng,nsg=nsg)
specgroups(x$distmat,groupvector,groupinfo)
```

stressvals *Stress values for different dimensions of Kruskal's MDS*

Description

Computes Kruskal's nonmetric multidimensional scaling isoMDS on alleleobject or prab-objects for different output dimensions in order to compare stress values.

Usage

```
stressvals(x,mdsdim=1:12,trace=FALSE)
```

Arguments

x	object of class alleleobject or link{prab}. generated by alleleinit or prabinit.
mdsdim	integer vector of MDS numbers of dimensions to be tried.
trace	logical. trace-argument for isoMDS (should trace information be printed during execution?).

Details

Note that zero distances between non-identical objects are replaced by the smallest nonzero distance divided by 10 to prevent isoMDS from producing an error.

Value

A list with components

MDSstress	vector of stress values.
mdsout	list of full outputs of isoMDS.

Author(s)

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Examples

```
  options(digits=4)
  data(tetragonula)
  set.seed(112233)
  taiselect <- sample(236,40)
# Use data subset to make execution faster.
  tnb <-
  coord2dist(coordmatrix=tetragonula.coord[taiselect,],
    cut=50,file.format="decimal2",neighbors=TRUE)
  ta <- alleleconvert(strmatrix=tetragonula[taiselect,])
```

```
tai <- alleleinit(allelematrix=ta,neighborhood=tnb$nblist)
stressvals(tai,mdsdim=1:3)$MDSstress
```

`tetragonula` *Microsatellite genetic data of Tetragonula bees*

Description

Genetic data for 236 Tetragonula (Apidae) bees from Australia and Southeast Asia, see Franck et al. (2004). The data give pairs of alleles (codominant markers) for 13 microsatellite loci.

Usage

```
data(tetragonula)
```

Format

Two objects are generated:

tetragonula A data frame with 236 observations and 13 string variables. Strings consist of six digits each. The format is derived from the data format used by the software GENEPOP (Rousset 2008). Alleles have a three digit code, so a value of `"258260"` on variable V10 means that on locus 10 the two alleles have codes 258 and 260. `"000"` refers to missing values.

tetragonula.coord a 236*2 matrix. Coordinates of locations of individuals in decimal format, i.e. the first number is latitude (negative values are South), with minutes and seconds converted to fractions. The second number is longitude (negative values are West).

Details

Reads from example data file `Heterotrigona_indoFO.dat`.

Source

Franck, P., E. Cameron, G. Good, J.-Y. Rasplus, and B. P. Oldroyd (2004) Nest architecture and genetic differentiation in a species complex of Australian stingless bees. *Mol. Ecol.* 13, 2317-2331.

Rousset, F. (2008) genepop'007: a complete re-implementation of the genepop software for Windows and Linux. *Molecular Ecology Resources* 8, 103-106.

Examples

```
data(tetragonula)
```

toprab *Convert abundance matrix into presence/absence matrix*

Description

Converts abundance matrix into binary (logical) presence/absence matrix (TRUE if abundance>0).

Usage

```
toprab(prabobj)
```

Arguments

prabobj	object of class prab.

Value

Logical matrix with same dimensions as prabobj$prab as described above.

Author(s)

Christian Hennig <christian.hennig@unibo.it> https://www.unibo.it/sitoweb/christian.hennig/en

Examples

```
data(siskiyou)
x <- prabinit(prabmatrix=siskiyou, neighborhood=siskiyou.nb,
          distance="none")
toprab(x)
```

unbuild.charmatrix *Internal: create allele list out of character matrix*

Description

Creates a list of lists, such as required by alleledist, from the charmatrix component of an alleleobject.

Usage

```
unbuild.charmatrix(charmatrix,n.individuals,n.variables)
```

Arguments

charmatrix	matrix of characters in which there are two rows for every individual corresponding to the two alleles in every locus (column). Entries are allele codes but missing values are coded as NA.
n.individuals	integer. Number of individuals.
n.variables	integer. Number of loci.

Value

A list of lists. In the "outer" list, there are n.variables lists, one for each locus. In the "inner" list, for every individual there is a vector of two codes (typically characters, see alleleinit) for the two alleles in that locus.

Author(s)

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See Also

alleleinit, build.charmatrix

Examples

```
data(tetragonula)
tnb <-
coord2dist(coordmatrix=tetragonula.coord[1:50,],cut=50,file.format="decimal2",neighbors=TRUE)
ta <- alleleconvert(strmatrix=tetragonula[1:50,])
tai <- alleleinit(allelematrix=ta,neighborhood=tnb$nblist,distance="none")
str(unbuild.charmatrix(tai$charmatrix,50,13))
```

---

veronica *Genetic AFLP data of Veronica plants*

Description

0-1 data indicating whether dominant markers are present for 583 different AFLP bands ranging from 61 to 454 bp of 207 plant individuals of Veronica (Pentasepalae) from the Iberian Peninsula and Morocco (Martinez-Ortega et al., 2004).

Usage

```
data(veronica)
```

Format

Two objects are generated:

veronica 0-1 matrix with 207 individuals (rows) and 583 AFLP bands (columns).

veronica.coord a 207*2 matrix. Coordinates of locations of individuals in decimal format, i.e. the first number is latitude (negative values are South), with minutes and seconds converted to fractions. The second number is longitude (negative values are West).

Details

Reads from example data files `MartinezOrtega04AFLP.dat`, `MartinezKoord.dat`.

Source

Martinez-Ortega, M. M., L. Delgado, D. C. Albach, J. A. Elena-Rossello, and E. Rico (2004). Species boundaries and phylogeographic patterns in cryptic taxa inferred from AFLP markers: Veronica subgen. Pentasepalae (Scrophulariaceae) in the Western Mediterranean.*Syst. Bot.* 29, 965-986.

Examples

```
data(veronica)
```

---

waterdist *Overwater distances between islands in the Aegean sea*

Description

Distance matrix of overwater distances in km between 34 islands in the Aegean sea.

Usage

```
data(waterdist)
```

Format

A symmetric 34*34 distance matrix.

Details

Reads from example data file `Waterdist.dat`, in which there is a 35th column and line with distances to Turkey mainland.

Source

B. Hausdorf and C. Hennig (2005) The influence of recent geography, palaeography and climate on the composition of the faune of the central Aegean Islands. *Biological Journal of the Linnean Society* 84, 785-795.

Examples

```
data(waterdist)
```

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